1 CD38 in the Age of COVID-19: A Medical Perspective

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66 **ABSTRACT** (164 words)

This medical review addresses the hypothesis that CD38/NADase is at the center of a functional 67 axis (i.e., intracellular Ca2+ mobilization/IFNy response/ROS burst) driven by SARS-CoV-2 infection, as 68 69 already verified in Respiratory Syncytial Virus pathology and CD38 activity in other cellular settings. Key 70 features of the hypothesis are that: i) the substrates of CD38 (e.g., NAD⁺ and NADP⁺) are depleted by viral-induced metabolic changes; ii) the products of the enzymatic activity of CD38 (e.g., 71 cADPR/ADPR/NAADP) and related enzymes (e.g., PARPs, Sirtuins, ADP-ribosyl hydrolase) are 72 involved in the anti-viral and proinflammatory response that favors the onset of lung immunopathology 73 74 (e.g., cytokine storm and organ fibrosis); and iii) the pathological changes induced by this kinetic mechanism may be reduced by distinct modulators of the CD38/NAD⁺ axis (e.g., CD38 blockers; NAD⁺ 75 suppliers, among others). This view is supported by arrays of associative basic and applied research 76 data which are herein discussed and integrated with conclusions reported by others in the field of 77 inflammatory, immune, tumor and viral diseases. 78

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80 CALL-OUT BOX

Although morbidity and mortality rates secondary to the inflammatory and systemic fibrotic conditions of COVID-19 patients are of great concern, only a very few specific drugs are available for treatment.

Emerging evidence supports the hypothesis that the CD38 ectoenzyme and products controlled by the CD38/NAD+ axis may play significant roles in the pathogenesis of the disease.

The use of CD38-targeted therapies may be a new and viable treatment option in life-threatening COVID-19.

81

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86 ABBREVIATIONS

- 87 (Not included in the List of Commonly Accepted Abbreviations)
- 88 ACE: Angiotensin-Converting Enzyme
- 89 ADPR: Adenosine diphosphate ribose
- 90 ADO: Adenosine
- 91 ARH: ADP-ribosylhydrolase
- 92 ARDS: Acute Respiratory Distress Syndrome
- 93 AhR: Aryl hydrocarbon Receptor
- 94 ATRA: All-trans retinoic acid
- 95 8-Br-cADPR: 8-Bromo-cADPR
- 96 cADPR: cyclic ADPR
- 97 Ca²⁺: ionic calcium
- 98 CD38: Cluster of Differentiation 38
- 99 COVID-19: Coronavirus disease 2019
- 100 CoV: Coronavirus
- 101 COPD: Chronic Obstructive Pulmonary Disease
- 102 CSS: Cytokine Storm Syndrome
- 103 DAMPs: Damage-Associated Molecular Patterns
- 104 DCs: Dendritic cells
- 105 Egr-1: Early growth response-1
- 106 ENPP1: ectonucleotide pyrophosphatase/phosphodiesterase 1
- 107 e5'NT: 5'ecto-nucleotidase
- 108 EL: Endolysosome
- 109 ER: Endoplasmic Reticulum
- 110 GCSF: Granulocyte-Colony Stimulating Factor
- 111 hMDDCs: human Monocyte-Derived Dendritic Cells
- 112 IFN-1: type 1 interferon
- 113 IRF: Interferon-Responsive Element
- 114 ISGs: Interferon-Stimulated Genes
- 115 mART: (mono) ADPribosyl transferase
- 116 MERS-CoV: Middle East Respiratory Syndrome Coronavirus
- 117 MDSC: Myeloid Derived Suppressor Cell
- 118 NA: Nicotinic acid
- 119 NAADP: Nicotinic acid adenine dinucleotide phosphate
- 120 NAM: Nicotinamide

- 121 NMN: Nicotinamide mononucleotide
- 122 NK cell: Natural Killer cell
- 123 NLRP3: NLR family pyrin domain containing protein 3
- 124 NRF2: Nuclear factor erythroid 2-related factor 2
- 125 NR: Nicotinamide riboside
- 126 nsp: non-structural protein
- 127 PAMPs: Pathogen-Associated Molecular Patterns
- 128 PM: Plasma Membrane
- 129 PARP: Poly (ADP-ribose) polymerase
- 130 RAS: Renin-Angiotensin System
- 131 RyR: Ryanodine Receptor
- 132 RSV: Respiratory Syncytial Virus
- 133 SARS-CoV-2: Severe Acute Respiratory Syndrome Coronavirus 2
- 134 SIRT: Sirtuin
- 135 ss-RNA: single strand-RNA
- 136 TLR: Toll-like-receptor
- 137 TMPRSS2: Transmembrane protease serine 2
- 138 TPC: Two-Pore Channel
- 139 Trp: Tryptophan
- 140 Treg: T regulatory lymphocyte
- 141
- 142

143 I. GENERAL PREMISE

144 A. Starting point

This perspective paper is grounded in observations from a 2018 study on the modulation of CD38 145 146 during Respiratory Syncitial Virus (RSV) infection in monocytes and macrophages (203). Upon 147 activation of the adaptive immune system during RSV infection, human monocyte-derived dendritic cells (hMDDCs) up-regulate CD38 expression and affect the ability to activate T cell proliferation (47). 148 Similarly, during other viral infections, overexpression of CD38 by both CD4⁺ and CD8⁺ T lymphocytes, 149 results in nicotinamide adenine dinucleotide (NAD⁺) depletion (201, 242). During viral infection, the 150 151 infiltration of monocyte-derived macrophages is accompanied by release of high levels of reactive oxygen species (ROS) and proinflammatory cytokines (164). In the case of RSV, the innate immune 152 response is initiated by recognition of single-stranded viral RNA (ssRNA) and secretion of type 1 153 interferon (IFN-1) by infected cells. 154

IFNs engage autocrine- or paracrine-specific receptors to induce expression of a set of IFN-155 156 stimulated genes (ISGs), which inhibit viral replication by reprogramming the cellular metabolism. Moreover, ISGs are inhibited by the anti-oxidant N-acetyl cysteine, further highlighting the role of ROS 157 in the process of anti-viral responses (46, 203). It is known that CD38 is involved in angiotensin (Ang) 158 II-induced intracellular Ca²⁺ release and ROS production (141). The ROS process in RSV-infected 159 hMDDCs is under the control of CD38 (203) and its catalytical activity is up-regulated as assessed by 160 measuring the accumulation of adenosine diphosphate-ribose (ADPR) after adding NAD⁺ as a substrate 161 (110, 203). This means that NAD⁺ consumption and generation of metabolic products by the enzymatic 162 functions of CD38 are involved in the induction of anti-viral and proinflammatory responses. 163

This paper seeks to identify the underlying basis of CD38 involvement in the response to Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) responsible for the coronavirus disease (COVID-19) pandemic (105). It does so by examining some of the key metabolic steps controlled by CD38 and its role in the immune response.

This paper seeks to identify the underlying basis of CD38 involvement in the response to Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) responsible for the coronavirus disease (COVID-19) pandemic (105). It does so by examining some of the key metabolic steps controlled by CD38 and its role in the immune response.

172 B. The COVID-19 disease

173 SARS-CoV-2 causes COVID-19, which, at time of this writing, has surpassed 100 million 174 confirmed cases and resulted in over 2% deaths recorded in more than 200 countries 175 (https://www.who.int/emergencies/diseases/novel-coronavirus-2019/situation-reports) (accessed on 176 February 2021).

177 C. The SARS-CoV-2 virus and cell entry

Pathogenesis of SARS-CoV-2 infection (Fig. 1A-C) starts when the trimeric viral spike (S) 178 179 glycoprotein binds to the human cell surface type I transmembrane angiotensin-converting enzyme 2 receptor (ACE2R), followed by proteolytic priming of the S protein. It contains two subunits: i) S1, which 180 181 has two major structural elements, the receptor-binding domain (RBD) and the N-terminal domain 182 (NTD), and ii) S2, which mediates virus-cell membrane fusion after the RBD engages ACE2 (243). A 183 two-step sequential protease cleavage model has been proposed for activation of S protein priming. A first cleavage between S1 and S2 activates a nick on S2' site, by host proteases: the cellular 184 transmembrane protease serine 2 (TMPRSS2) and furin, respectively (106, 118). Besides TRPRSS, 185 186 other proteases have also been implicated in facilitating virus entry. Indeed, the extracellular protease plasmin is also able to nick the spike at the S1/S2, a furin cleavage site that increases its ability to bind 187 with ACE2R of host cells (13). 188

Once the endocytic uptake is unlocked, the viruses uncoat the genome and release the genetic 189 material, namely ss-RNA, to initiate replication. The coronavirus (CoV) genome does not encode for 190 enzymes necessary for the synthesis of proteins, amino acids, lipids or nucleotides. Therefore, SARS-191 CoV-2 exploits the host cell for its own replication and to protect its ss-RNA from anti-viral immunity 192 (261). To ensures its integrity, viral RNA is capped and methylated at the 5'end by CoV-non-structural 193 protein (nsp) (e.g., methyltransferase, MTase) (245), thereby resembling host mRNA to promote 194 195 translation and to prevent its degradation. All the successive events occur in the nucleus and cytoplasm 196 (66).

ACE2 was originally identified as the receptor of other SARS CoVs (133) as well as of the RSV (203). Of note, ACE2 is also a metallocarboxypeptidase enzyme, which catalyzes the conversion of the substrate angiotensin (Ang)-II to Ang-1–7 in the Renin-Angiotensin System (RAS) (75), as shown in Fig.1B. Besides ACE2, it has also been suggested that CD26, the host receptor for MERS-CoV (239), and CD147 (120), serve as endocytic cell entry for SARS-CoV-2.

The ACE2R is expressed by endothelial and epithelial cells present in different organs, such as lungs, heart, gut, kidneys, brain, and placenta which are all susceptible to viral infection (226, 246, 250). In the lungs, ACE2 is expressed by cells of the upper or lower respiratory tract, a critical step for initiation and clinical presentation of the viral infection. Both SARS-CoV-2 and RSV mainly affect the lower respiratory tract (Fig.1C). Pathognomonic signs generally found in human diseases and caused by respiratory virus (e.g., RSV and SARS-CoV-2), are involved in an hyperimmune response causing lung pathology (203, 250).



Figure 1. Schematic illustration of the SARS-CoV-2 molecular structure and essential 210 mechanisms of viral infection and outcomes. A) The SARS-CoV-2 genome encodes non-structural 211 proteins (nsp1-nsp16) (not shown) and four structural proteins: spike (S) glycoprotein, envelope, 212 membrane, and nucleocapsid phosphoprotein, which together ensure replication of the virus in the host 213 cell. B) The octapeptide Ang II is originated from the decapeptide Ang I by soluble ACE2 enzymatic 214 activity. Ang II acts via AT1Rs while Ang (1-7), generated from Ang II by ACE2 carboxypeptidase, acts 215 via the Mas receptor (MasR). SARS-CoV-2 binding to the ACE2 catalytic receptor (ACE2R) enhances 216 lung inflammation by reducing ACE2 activity and increasing Ang II. Depletion of ACE2 activity decreases 217 the production of Ang 1-7, which has an anti-inflammatory and anti-fibrotic activity. C) SARS-CoV-2 and 218 RSV preferentially bind to the ACE2R expressed by alveolar epithelial cells and macrophages in the 219 lower human respiratory tract. 220

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II. CD38 CHARACTERISTICS AND FUNCTIONS POTENTIALLY LINKED TO THE HOST RESPONSE TO SARS-CoV-2 INFECTION.

224 A. CD38

225 CD38 is a multifunctional cell protein endowed with signaling receptor and enzymatic features 226 and was initially identified as a lymphocyte antigen by monoclonal antibody typing (195). CD38, present 227 outside of the cell (45) and also intracellularly in the nucleus and organelles (1), is associated with 228 important diseases, such as AIDS, autism, diabetes, chronic lymphocytic leukemia, and multiple 229 myeloma (Table 1). These characteristics of CD38 have been comprehensively reviewed (158).

230 1. CD38 as an enzyme

CD38 is a 43.7-kDa transmembrane glycoprotein, which also exists in a 39-kDa soluble form that retains its biochemical features in both normal and pathological fluids (80, 146). Recognition of structural and functional similarities between human CD38 and the enzyme ADP-ribosyl cyclase, purified from the sea mollusk *Aplysia*, allowed attribution of enzymatic activities to CD38 (137). Indeed, at physiological pH, CD38 catalyzes several enzymatic reactions: i) the conversion of nicotinamide adenine dinucleotide (NAD⁺) to adenosine diphosphate ribose (ADPR) (NAD⁺- glycohydrolase activity);

ii) the conversion of NAD⁺ to cyclic ADPR (cADPR) (cyclase activity), and iii) the hydrolysis of cADPR 237 to ADPR (hydrolase activity). At acidic pH, CD38 runs iv) the conversion of NADP⁺, the phosphorylated 238 equivalent of NAD⁺, to nicotinic acid adenine dinucleotide phosphate (NAADP) (NAADP-synthase 239 240 activity) in the presence of nicotinic acid (NA) and the degradation of NAADP to ADPR.P (NAADP-241 hydrolase activity) (Fig. 2). All of the reaction products are second messengers involved in the regulation of cytoplasmic Ca²⁺ fluxes (139). NAD⁺-glycohydrolase, the main enzymatic activity of CD38, is not 242 modified in the presence of anti-CD38 human or murine antibodies. On the contrary, cyclase activity is 243 highly inhibited, while hydrolase activity is mildly activated (11, 112, 113). This data provides further 244 support for considering extracellular CD38 primarily as a NAD⁺-glycohydrolase (107). CD38 is also able 245 to catalyze the degradation of intracellular NAD⁺ precursors [(e.g., nicotinamide mononucleotide (NMN) 246 and nicotinamide (NAM)] (32, 109). 247





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Figure 2. CD38 enzymatic activities. CD38 catalyzes several enzymatic reactions: at neutral pH i) the 250 conversion of nicotinamide adenine dinucleotide (NAD⁺) into adenosine diphosphate ribose (ADPR) 251 (NAD⁺-glycohydrolase activity); ii) the conversion of NAD⁺ into cyclic ADPR (cADPR) (cyclase activity); 252 iii) the hydrolysis of cADPR into ADPR (hydrolase activity). At acidic pH, iv) the conversion of NADP⁺, 253 254 the phosphorylated equivalent of NAD⁺, into nicotinic acid adenine dinucleotide phosphate (NAADP) (NAADP-synthase activity) in the presence of nicotinic acid (NA) and the degradation of NAADP into 255 ADPR.P (NAADP-hydrolase activity). All of the reaction products are second messengers involved in 256 the regulation of cytoplasmic Ca²⁺ fluxes and the generation of immunosuppressive adenosine (see text 257 and Fig. 3) 258

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260 *CD38* gene ablation experiments provide strong evidence that the enzymatic activity of CD38 is

responsible for producing cADPR and NAADP, because formation of both nucleotide messengers is

abrogated when the CD38 gene is deleted, indicating that CD38 is the dominant enzyme responsible 262 for their synthesis (139). The nucleotide messengers regulate diverse cell functions by mobilizing 263 intracellular Ca²⁺ stores: i) NAADP-elicited Ca²⁺ release, important for SARS-CoV-2 entry into cells 264 (189), from the two-pore channels (TPCs), situated in acidic endolysosomes (EL) (24); ii) cADPR 265 enhances Ca²⁺ release via the activation of the ryanodine receptor (RyR) (158) situated in the membrane 266 of the endoplasmic reticulum (ER). cADPR can also activate the Ca²⁺ influx channel Transient Receptor 267 Potential Melastatin 2 (TRPM2) at the cell plasma membrane (PM), in synergy with ADPR (135) (Fig. 268 3). Although physically separated, the Ca^{2+} stores in the ER and the EL can interact: in fact, Ca^{2+} 269 released from the EL stores can be sequestered by the ER stores, boosting the latter for enhanced 270 release of Ca²⁺ through RyRs by cADPR (Fig. 3). 271

Further evidence for this view is that stimulation of the T cell receptor-CD3 complex results in rapid NAADP formation in response to a stimulus. By contrast, an increase in cADPR concentration is delayed, which indicates that NAADP serves as a second messenger initiating role in T cell Ca²⁺ signaling (35, 100).

It was initially thought that CD38 operated exclusively in the extracellular compartment 276 containing the physiological substrates, with the products of the catalytic reaction being used inside the 277 cell, creating a sort of topological paradox. Most immune and non-immune cells express CD38 on the 278 surface, with the catalytic domain exposed to the outside. These are referred to as type II CD38. 279 Extracellular NAD⁺ and NADP⁺ substrates are metabolized by type II CD38 into cADPR/ADPR and 280 281 NAADP, respectively, acting in an autocrine mode for signaling (45). Substrates share the mechanism 282 for extrusion either by cell lysis under pathological conditions (e.g., inflammation or oncogenesis) or by 283 transportation through the connexin 43 (Cx43) hemichannels (21). Extracellular metabolites are able to 284 reenter the cell using concentrative nucleoside transporters (CNTs), where they can also acts in a paracrine mode on neighboring cells (76). Type II CD38 is also compartmentalized in the EL/RE 285 organelles (251). Accordingly, the intracellular exploitation of type II CD38 metabolites targeting 286 intracellular Ca²⁺ release machineries, give rise to this topological enigma, only recently partially 287 disentangle (140). 288

These studies demonstrated the existence of a CD38 protein (referred to as type III CD38), 289 290 whose catalytic domain faces the intracellular compartment. This functionally-active molecule is 291 expressed on the inner cell membrane and in the ER and produces intracellular cADPR with high 292 efficiency (145). Type III CD38 is a non-glycosylated protein and thus devoid - in contrast to type II-CD38 - of disulfide bridges (140), but whose formation during folding allows cADPR generation. The 293 autocrine/paracrine mechanisms of type II- and type III-CD38 work in concert to harmonize the 294 paradoxical regulatory issue. In mechanistic terms, and in line with previous observations on the pH 295 dependency of CD38, the resulting cADPR and ADPR products are synthesized at neutral pH, while 296

NAADP is synthesized at acidic pH (100, 138). The EL is highly acidic – and therefore not favorable for the cyclase activity of CD38 – thus pointing to EL the cellular compartment for the biogenesis of NAADP (Fig. 3). Extracellular NAADP can also be transported into the cell cytoplasm, where NAADP, either from inflow or *in situ* generated, is delivered to the EL to induce Ca²⁺ release from stores in response to various physiological stimuli (81). Additional findings were that cells expressing type III CD38 had the highest cADPR levels after induction by cytokines, and thus may be directly responsible for producing intracellular cADPR (258), targeting RyRs in the ER (Fig. 3).





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Figure 3. Schematic illustration of intracellular signaling mediated by the CD38/NAD⁺ axis. A) 306 The NADPase and NADase enzymes are responsible for the formation of the Ca²⁺-releasing 307 messengers through the use of phosphorylated (NADP⁺) or non-phosphorylated NAD+, respectively. 308 Second messengers generated as products are: NAADP, cADPR, and ADPR. NAADP-elicited Ca²⁺ is 309 released from the two-pore channel (TPC) receptor situated in acidic endolysosomes (EL), and cADPR 310 serves as the trigger and booster for Ca2+ release via the activation of the ryanodine receptor (RyR), 311 situated in the endoplasmic reticulum (ER). ADPR elicits Ca²⁺ influx through the transient receptor 312 melastatin 2 (TRPM2) situated in the plasma membrane (PM). B) ADPR can also be sequentially 313 metabolized by ectonucleotidases (CD203a/ectonucleotide pyrophosphatase/phosphodiesterase 1 314 315 (ENPP1) and CD73/5'-ectonucleotidase (5'eNT) for the formation of extracellular adenosine (ADO).

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317 2. CD38 as a receptor

Functional and structural data indicate that the promoter region of the human *CD38* gene, located at chromosome 4, is regulated by several nuclear factors including RARα (retinoic acid receptor), RARE (retinoic acid-responsive element), GREs (glucocorticoid-responsive element), IRF

(interferon-responsive element) and NF-kB (72). Further, CD38 expression is induced and regulated by 321 322 several soluble factors including cytokines and chemokines (5). The role of CD38 as a receptor was 323 confirmed by co-modulation experiments, indicating that the molecule displays lateral associations with other molecules sharing signal pathways. In this way, CD38 overcomes the steric hindrance of its very 324 325 short cytoplasmic tail by interacting symbiotically with skilled receptors on different immune cells (e.g., 326 T lymphocytes, NK cells and monocytes) (158). The CD38 protein, assembled as a transmembrane receptor, influences both innate and adaptive immune responses by regulating the trafficking of cells 327 (e.g., macrophages, dendritic cells, lymphocytes, and neutrophils) to the sites of inflammation (77). For 328 329 migration purposes, CD38 expresses two hyaluronate-binding sites in the extracellular domain (183) 330 and thus interacts with its counterreceptor CD31 (PECAM, platelet/endothelial cell adhesion molecule-1) (48, 180). CD38 is also related to T helper type 1 polarization and dendritic cells (DCs) chemotaxis 331 (77, 190). 332

333 B. The CD38 catalytic receptor and inflammation

In addition to being a surface cell differentiation and activation marker, it was later observed that 334 335 CD38 can induce the release of different cytokines after specific agonistic monoclonal antibody (mAb) ligation (5). Conversely, it was observed that IFN-y alone induces CD38 in human 336 macrophage/monocytes (177) and that Vitamin D causes myeloid cells to express surface markers of 337 monocytic cell differentiation (e.g., CD38, CD14, CD11b), converting macrophages into potent 338 immunosuppressive cells (191). CD38 activity in human macrophages is prevalently detected 339 340 intracellularly, with its primary function being i) the performance of cADPR and NAADP for Ca²⁺ 341 regulation; ii) the contribution to inflammatory cytokine secretion, and iii) cooperation in reprogrammed metabolic adaptations (e.g., increased glycolytic activity). Taken together, these data are consistent with 342 the role of inflammatory marker for human macrophage/monocyte CD38 in inflammatory processes (3). 343

Type 1 interferons (IFN α /IFN β) as well as other factors such as the RAS component Ang II (via activation of NF-kB) up-regulate CD38 expression (28, 162) in pro-inflammatory cytotoxic human M1 polarized macrophages, but not anti-inflammatory M2 (41, 117, 214). In turn, signaling through NF-kB, likely the primary transcription factor involved in the appearance of most of the proinflammatory genes, is amplified by CD38 (122). Further, CD38 overexpression promotes a glycolytic adaptation in human macrophages (117).

Related to this metabolic event, nucleotides (such as NAD⁺ and ATP) are released during the early phase of viral inflammation, acting as danger signals that alert the immune system through binding to P2 type of purinergic receptors (P2Rs) (22, 115). Consumed nucleotides are re-builded by enzymatic salvage pathways to restore extracellular homeostasis. As shown in Fig. 4, NAD⁺ scavengers are nucleotide-catabolizing ectoenzymes (e.g., CD38/NADase, ectonucleoside pyrophosphatase phosphorylase, ENPP1/CD203a, and 5'-ectonucleotidase, 5'eNT/CD73) that generate adenosine (ADO) as end product, which can re-enter the cell to reconstitute the pool of purine nucleotides (110,
158). Alternatively, extracellular ADO, and probably inosine (INO), activates purinergic P1 receptors
(P1Rs) to dampen excessive inflammation, thus opposing inflammatory functions of P2R signaling (186).

Nucleotides released during viral inflammation also exert immunoregulatory roles *in vivo* (32, 203). For instance, macrophages not only express ACE2, but also high levels of CD38, the main consumer of human NAD⁺. A reasonable model is that in hyperstimulated macrophages, the NLR family pyrin domain containing protein 3 (NLRP3) inflammasome can be directly activated by SARS-CoV-2 via a CD38-mediated Ca²⁺-dependent mechanism. This was posited during 2015 SARS epidemic (181).

A preliminary conclusion is that CD38 is involved in specific steps of viral inflammation by modulating immune response and by regulating Ca^{2+} signaling in different cell populations and tissues.

366 C. The CD38 catalytic receptor and immune response

The immune system exploits cell and humoral responses to attack viruses. Of all the various steps in COVID-19 immunity (237), here we analyze those potentially linked to CD38.

In SARS-CoV-2, innate immune response is activated when macrophages encounter viral 369 pathogen-associated molecular patterns (PAMPs) from invading SARS-CoV-2 ss-RNA. PAMPs up-370 371 regulates CD38 and activates innate immune pathways through Toll-like receptors (TLRs) and NLRP3 372 inflammasome activation (240). Downstream signaling drives the secretion of a range of proinflammatory cytokines, including IL1- β , IL1RA, IL-6, IL-7, IL-18, IL-10, IFNy, and TNF α (61). This 373 374 leads to rapid recruitment of monocyte/macrophages to the lung in early phases of infection. The successive production of IFN α /IFN β limits propagation of the virus (142). At the same time, IFNs 375 modulate the adaptive immune responses by increasing the expression of anti-viral specific genes 376 377 (ISGs) in neighboring cells (198).

378 For their part, CoVs escape immune responses by provoking a disbalance between anti-viral and proinflammatory responses. It is hypothesized that SARS-CoV-2 exploit the up-regulation of host 379 380 ACE2R, by increased expression of ISGs and CD38 in human lung epithelial cells, to enhance viral 381 infection (141, 260). Other steps facilitating SARS-CoV-2 infection involve the suppression of IFN expression by inhibiting the host sensor machinery or its downstream signaling (240). Other cell types, 382 including endothelial cells, are indirectly activated by circulating IL-6 and soluble IL-6 receptor 383 complexes, with massive cytokine production and cell apoptosis (227). Apoptotic infected endothelial 384 and epithelial cells contribute to tissue inflammation by releasing damage-associated molecular patterns 385 (DAMPs) into the extracellular environment and IL-1 β upon NLRP3 inflammasome activation. The 386 overproduction of IL-1β then activates macrophages, NK and T cells, amplifying inflammation and 387 388 facilitating tissue infiltration through the up-regulation of adhesion molecules by lung endothelial cells. 389 Indeed, IL-1 β increases hyaluronan synthetase levels, and consequently matrix hyaluronate (12), which 390 is reported as an adhesive ligand of CD38 (183).

391 D. The CD38 catalytic receptor in cell adhesion and thrombosis

392 Up-regulation of molecules involved in cell adhesion (i.e., CD38 and CD31) has two-fold 393 consequences: lymphopenia and thrombosis, which are both predictors of COVID-19 disease severity 394 (224, 230).

Viral infection inducing excessive antigenic stimulation may cause a drastic decay in circulating immune cells with progressive T cell anergy or exhaustion (50, 143). Mechanisms leading to lymphopenia can be due to (i) a direct effect on lymphocytes or indirect action destroying lymphatic organs; or (ii) a disordered inflammatory cytokine reaction leading to lymphocyte apoptosis (144). CD38/NADase might be directly involved in these events. Indeed, CD38 activation by increased Ang II levels may intensify NAD⁺ depletion. In turn, this condition affects NAD⁺-dependent enzymes (e.g., Sirtuins and PARPs), which are known regulators of cell viability and death (107).

CD38 express by immune cells is reported as interacting with extracellular matrix hyaluronate 402 and with CD31 (48). The balance between dissemination of immune cells (CD38^{high+} cells) to peripheral 403 blood and tissue retention in the respiratory tract is the result of the interplay between these molecules. 404 Indeed, CD38^{high+} promotes cell attachment to hyaluronate, whereas the interaction of CD38 with CD31 405 on endothelial cells results in retention prevalently in tissues (in the lungs) and a weaker egress of 406 immune cells to the peripheral blood, as has been shown in a leukemia model (82). Therefore, the 407 408 trapping of exhausted T lymphocytes in the lungs may contribute to lymphopenia. Lastly, lymphocyte 409 functions may be impaired by products derived from metabolic disorders, such as lactic acidemia (121) 410 (vide infra).

411 COVID-19 comorbidities feature elevated levels of the extracellular plasmin, a protease involved in degradation of the fibrin matrix formed by the activity of thrombin during the process of thrombosis 412 (119, 224). Further, dysregulated Ang II due to loss of ACE2R by SARS-CoV-2, results in increased 413 signaling through i) purinergic P2R(s), and by ii) the serine protease thrombin, leading both to platelet 414 activation and thrombosis, emerging features of COVID-19 (224). Thrombin induces platelets activation 415 via mobilization of intracellular Ca²⁺, a process mediated by CD38 metabolic products, cADPR and 416 NAADP (176). Moreover, as a platelet agonist, thrombin stimulates the association of CD38 enzymatic 417 activities with the platelet cytoskeleton (236). As said, inflammatory conditions observed in COVID-19 418 419 are associated with the extracellular release of nucleotides, acting as ligands of purinergic receptors (224). P2Rs signaling is a key mechanism for platelet activation, which contributes to 420 thromboinflammation and fibrosis (68). Indeed, an inflammatory P2R-associated release of IL-8 and 421 elastase from neutrophils contributes to the pathogenesis of chronic obstructive pulmonary lung disease 422 (COPD) (115). This finding suggest that nucleotide-activation of P2Rs can lead to inflammation, tissue 423 fibrosis, as well as to a NAD⁺-dependent Sirtuins inhibition associated to ROS production (68). 424 Purinergic and thrombotic-mechanisms can synergistically be activated during thrombosis (60). 425

Therefore, antagonistic drugs that target thrombin or P2Rs, may provide a useful therapy to blunt inflammatory diseases, such as COPD and COVID-19 (224). Similarly, catalyzing the conversion of ATP/NAD⁺ to ADO, thus terminating P2R effects, are already exploited in the treatment of inflammatory conditions in human patients (169).

Consequently, CD38 may serve as a molecular target for i) immune cell trapping in the lungs
 and ii) monitoring a down-modulation of macrophages during viral respiratory diseases (203). Further,
 it may help track iii) lymphopenia and thrombosis resulting from uncontrolled activation of immune cells.

433 E. The CD38 catalytic receptor and immuno-metabolic adaptations

434 1. The NAD⁺ metabolome

The metabolism of NAD⁺ (NAD⁺ metabolome) is involved in a variety of normal biological processes (182). As a tunable component of innate immunity, the NAD⁺ metabolome has become a target for therapeutic modulation of the NAD⁺ status, which potentially curbs viral infection (131). Observations support the view that the immune response to viral infections is linked to the NAD⁺ metabolome of the infected cells, as reported in Herpes virus and HIV-1 (93, 175).

NAD⁺ operates both intra-and extracellularly (Fig. 4). Extracellularly, NAD⁺ elicits signals acting 440 as a cytokine or serves as the substrate for a chain of nucleotidases led by CD38 to convert it to ADO, 441 a nucleoside involved in the control of inflammation and immune responses (110, 168). The extracellular 442 conversion of NAD⁺ varies significantly according to the tissue environment or health conditions. Indeed, 443 pathological settings are characterized by the NAD⁺ metabolome acting as a target of multiple 444 445 immunometabolic adaptations, as confirmed by a dysregulated NAD⁺ gene system upon in vivo SARS-446 CoV-2 infection (103). Analyzes of RNAseg data involved comparison with a gene set representative of 447 the NAD⁺ transcriptome coding for the enzymes responsible for i) NAD⁺ biosynthesis; ii) NAD⁺ phosphorylation to NADP⁺ and, iii) NAD⁺ consumption. First, primary cells infected by SARS-CoV-2 448 feature >3-fold depression of cellular NAD⁺ and NADP⁺, as compared to control cells. CD38 (and its 449 paralogue CD157) (158) are overexpressed (>2.5- and >1.5-fold, respectively) by infected human lung. 450 CD38 up-regulation and NAD⁺ depletion are paralleled by activation of the IFN-induced 451 452 mARTransferase (mART) (65). The SARS-CoV-2 genome encodes for nsp (245): among them, an ADPribosylhydrolase (ARH), an enzyme required for virulence that removes ADPR from proteins ribosylated 453 454 by mART (Fig.4). (2, 128).

The NAD⁺ metabolome is linked to the RAS/ACE2 system. On one side, NAD⁺ biosynthesis is regulated by the nutritional supply of NAD⁺ precursors through *de novo* pathway, which uses tryptophan (Trp), and the salvage pathway, which uses NAM/NA/NR (all referred to as vitamin B3), as primary sources (Fig. 4) (109). Trp catabolism in chronic viral infections reduces circulating levels of NAD⁺, resulting in exacerbated inflammation and low CD4⁺ T-cell recovery (163). On the other, the RAS system exerts a protective role in acute lung injury via ACE2 and by modulating Trp levels in peripheral blood (133). Indeed, in aminoacidic malnutrition, increasing Trp and vitamin B3 sources restores ACE2 activity
and prevents worsening of inflammation (102). In conclusion, epigenetic and pharmacological evidence
link the NAD⁺ metabolome to the RAS/ACE2 system. CD38 is activated by Ang II after ACE2 viral
blocking (Fig. 1B) and once the human lungs are infected, the virus may even try to suppress NAD⁺
production by the cells (16). NAD⁺ depletion leads suppression of both mitochondrial NAD⁺-dependent
signaling and resolution of inflammation (165).

Metabolomic studies indicate that under non-redox conditions NAD⁺ is mainly consumed by 467 CD38/NAD⁺-glycohydrolase, NAD⁺-dependent Sirtuins and -PARPs (Fig. 4) (182). Because of such 468 469 continuous NAD⁺ enzymatic degradation, its metabolite NAM and the other amidated and deamidated 470 NAD⁺ sources (e.g., NR, NMN, NA) needed for resynthesis of NAD⁺ are perforce scavenged (107). In fact, the anti-viral host defenses mounted by NAD⁺-dependent-PARPs and -Sirtuins are removed by 471 depleting the cell of NAD⁺ (107, 203). This is supported by mice models showing that increased NAD⁺ 472 levels augment the enzymatic activity of PARPs and Sirtuins, hindering CoV from hijacking the host 473 474 cellular machinery for replication (67).

475 CD38 is a crucial regulator of Sirtuins which modulate normal and pathological energy metabolism (124). Sirtuins are dependent on NAD⁺ biogenesis, and thus regulated by Trp or by 476 nicotinamide phosphatidyltransferase (NAMPT), the rate-limiting enzyme that converts NAM into NAD⁺ 477 478 (Fig. 4). Sirtuins and NAMPT participate in macrophage antiviral activity (43). In addition, CD38 activates the Sirtuin/NFkB pathway in a NAD⁺-dependent manner, since CD38 blocking increases NAD⁺ levels 479 480 and Sirtuin-1 activity in the nuclear, cytoplasmic and mitochondrial compartments (1, 34). The 481 pharmacological inhibition of NAMPT and Sirtuins, components of the macrophage IFN anti-viral 482 cascade, promotes growth of cytomegalovirus in both fibroblasts and macrophages (43). The central 483 role of the NAD⁺ metabolome in these cells is further supported by the notion that extracellular NAMPT behaves as a DAMP (159), which is elevated in COVID-19 patients with comorbidities (213). 484

The NAD⁺-consuming enzyme PARPs and the aryl hydrocarbon receptor (AhR) are 485 overexpressed in COVID-19 pathophysiology, and in other lung conditions (RSV and COPD) (17, 96). 486 487 Endogenous AhR ligands include Trp metabolite quinolinic acid in the de novo pathway and NA and NAM in the salvage pathway of NAD⁺ biogenesis (Fig. 4). As a transcription factor, AhR is involved in 488 489 microbial defense, cell proliferation, immunity and NAD⁺ metabolism (17). AhR targets NAD⁺ metabolome functional elements such as CD38 and PARPs that are regulating glucose and lipid 490 metabolism via Sirtuins. Deregulation of these pathways may facilitate COVID-19 and age-dependent 491 pathologies (87). Indeed, a proinflammatory milieu leads to up-regulation of the AhR which in turn 492 activates PARPs. Mucin overproduction by lung epithelial cells triggered by IFN-signaling thickens the 493 494 blood-air barrier and leads to hypoxia (150). Because mucin up-regulation is driven by AhR, this factor

involved in NAD⁺ homeostasis in cooperation with CD38, PARPs and Sirtuins, is a potential target for
the treatment of hypoxia in COVID-19 patients (114).

Overexpression of CD38 and PARPs in COVID-19 causes cell death mainly by depletion of NAD⁺
 (6). NAD⁺ boost improves blood flow and vascular vitality by promoting Sirtuins dependent increase of
 the levels of hydrogen sulfide (H2S), an endothelial signal regulator of NAD⁺ levels (44, 252). Since H2S
 intracellular activity ensures vascular repair after injury, the relevance of the integrity of the NAD⁺
 metabolome should be considered in an eventual SARS-CoV-2 infection of endothelial cells, known to
 express ACE2 receptors (70).

503 Oral administration of amidated NAD⁺ precursors (NR, NAM, and NMN) has been demonstrated 504 to be the most effective approach to replenishing NAD⁺ levels *in vivo*. Of these NAD⁺ precursors, NR 505 has been shown to have anti-inflammatory effects in different disease conditions in both preclinical and 506 clinical settings (55). Currently, a clinical trial of NR as a therapeutic option in COVID-19 patients is 507 ongoing (248).

NAM is a potent PARPs inhibitor that boost NAD⁺/NADP⁺ synthesis. Hence NAM reverses lung 508 injury caused by ischaemia, inhibits proinflammatory cytokines and is effective against HIV-1 infection 509 (87, 213). Another aspect of NAM effects that is relevant to the metabolome rewiring of NAD⁺ is their 510 contribution to maintaining homeostasis through the involvement of gut microbiota in NAD⁺ biogenesis 511 512 (53). NAM suppliers (such as NR and NMN) are thus potential candidates for use in COVID-19 treatment by replenishing NAD⁺ levels (6, 32, 55). NMN plays an anti-inflammatory role in preclinical models 513 514 decreasing the levels of lactic acidosis and IL-6. By reducing IL-6, NMN improves shock-induced 515 hyperglycemia, reducing inflammation (32).

All of the evidence seems to confirm that key events of the biosynthesis and consumption of NAD⁺ play significant roles in the anti-viral immune response. Consequently, NAD⁺ refueling by modulating the biosynthetic pathways or – alternatively - by reducing NAD⁺ consumption (34) may be of help in controlling the hyperimmune response to SARS-CoV-2 infection.



521

Figure 4. Pathways for NAD⁺ biogenesis and consumption. Intracellular NAD⁺ is synthesized either 522 from tryptophan (de novo pathway) or from nicotinamide riboside (NR), nicotinamide (NAM), or nicotinic 523 acid (NA) (salvage pathways). Once internalized, NAM and NR merge at the step of nicotinamide 524 mononucleotide (NMN), which is converted into NAD⁺, NA is converted to NA adenine dinucleotide 525 (NAAD), and then to NAD⁺. Depletion of NAD⁺ is associated with enzymatic reactions that take place 526 intracellularly: CD38/NAD⁺-glycohydrolase, PARPs and Sirtuins. NAD⁺ is also used as a cofactor by S-527 528 adenosylmethionine (SAM) for i) the generation of intracellular adenosine from methionine, and ii) the activity of a viral SAM-dependent Methyl Transferase (MTase) enzyme, composed by the SARS-CoV-529 2 non-structural proteins (nsp) 14 and 16, active for viral cap formation during viral replication. 530 531 Extracellular NAD⁺ is metabolized by CD38, the first enzyme within a purinergic signaling cascade that, together with CD203 and CD73, generates exogenous adenosine. 532

533

534 2. Alternative NAD⁺- consuming enzymes

CD38 consumes NAD⁺ in multiple ways, such as by i) mobilizing extracellular or intracellular 535 NAD^+ pools, depending on its membrane topological conformation (140), and by ii) degrading 536 extracellular NAD⁺ to generate NAM, which can cross the plasma membrane and be converted to NMN 537 and NAD⁺ through NAMPT and NMNAT (25, 109). Although CD38 is the major ectoenzyme responsible 538 for NAD⁺ metabolization in mammalian tissues (158), there is evidence for cADPR and NAADP 539 540 generation by other molecules. Indeed, depletion of extracellular NAD⁺ also occurs through the highly conserved CD38 homolog CD157/Bst1, a molecule which, however, exhibits very low NAD⁺-consuming 541 activity (158). Another ectoenzyme that degrades extracellular NAD⁺ is CD73/e5'NT, which successively 542 metabolizes NAD⁺ to NMN, and further to NR (84), to support intracellular NAD⁺ biosynthesis. In 543 particular, the cADPR levels in the brain of CD38-KO mice are consistent (190), indicating the existence 544

of a cADPR-synthesizing enzyme. This enzyme was identified in the brain as SARM1 (sterile alpha and 545 Toll/interleukin receptor motif-containing protein 1) (59, 153), which features NAD⁺-cyclizing activity 546 much higher than CD38, already known by its low (2%) cADPR yield after NAD⁺ dismantling activity 547 548 (138). The SARM1 molecule has no sequence similarity, but has the same cytosolic orientation as type 549 III CD38, and is able to catalyze the same set of NAD⁺-depleting multi-reactions after being activated by 550 endogenous NMN (25, 59, 259). CD38 is the main enzyme involved in the degradation of NMN in vivo (25). As an NMNase, CD38 controls the paracrine availability of extracellular NMN (but not NR or NA) 551 (32), and thus influences the accessibility of NMN to SARM1. 552

553 For extracellular signaling activities in immune cells, NAD⁺ uses purinergic P1 and P2 receptors and metabolizing ectoenzymes (CD38, CD203a and CD73) (110). Notably, recent data showed that 554 CD203a/ENPP1 also metabolizes 2',3' cyclic GMP-AMP dinucleotide (cGAMP), generating AMP and 555 GMP (7), all acting as modulators of immunity (123). Indeed, DNA/RNA released in the cytoplasm during 556 viral infection activates a cyclic GMP-AMP synthase (cGAS), forming cGAMP from cAMP/cGMP. 557 Interesting, cGAMP is an activator of STING (stimulator of interferon genes), that integrates together 558 with SARM1, a subset of Toll-Interleukin receptor (TIR) domain-containing proteins. Both proteins can 559 degrade NAD⁺ by acting as NAD⁺-hydrolases producing ADPR and NAM, thus supporting TIR domain-560 mediated sensing of innate immunity (152). Links between the cGAMP-STING pathway with CD203a 561 and NAD⁺ have emerged whereby the hydrolysis of cGAMP by CD203a attenuates cGAS-STING 562 signaling and, therefore, the depletion of NAD⁺ (187). Consequently, inhibitors of CD203a (207) could 563 564 help to combat viral activity by inhibiting cGAMP degradation and extracellular NAD⁺ consumption.

565 New aspects of NAADP generation were reported indicating that the CD38-base exchange reaction is not the enzyme responsible for in vivo generation of this nucleotide in human myometrial 566 cells (219). Of note, NAADP-dependent generation and the release of Ca²⁺ was experimentally 567 evidenced at physiological pH in response to histamine and oxytocin as modulators and with the use of 568 pharmacological inhibitors. On the other hand, an insulin sensitization by NAADP was reported to be 569 570 produced through both CD38-dependent and CD38-independent pathways (221). CD38 is still the only 571 molecule fully characterized as consuming NAD⁺ and synthesizing messengers (cADPR, ADPR and NAADP) in a variety of cells (158) (and references herein). 572

- A closer look at NAD⁺-consuming enzymes therefore reveals differences in chemical structure, tissue distribution, compartmentalization, metabolism, substrate affinities and response to specific modulators, which might affect the performance among redundant enzymes. A sensitive proxy for hierarchical selectivity among NAD⁺-consuming enzymes would be the level of physiological effects of each enzyme in different tissues and their different effects in clinical trial outcomes.
- 578 *3. Metabolic acidosis and adenosinergic activities*

Cell homeostasis depends on adenine nucleotides (e.g., NAD⁺/NADP⁺, and ATP). They produce 579 580 energy on the one hand, and generate anabolic products and second signal messengers (109) on the other. Energy production takes place in the cytoplasm, and glucose is transformed into pyruvate. Under 581 582 normal oxygenation, pyruvate enters the mitochondria, where it undergoes enzymatic processes 583 generating large quantities of ATP. In hypoxic cells (e.g., inflammation, tumors), pyruvate cannot enter 584 the mitochondria, but is converted to lactic acid. This step is marked by the generation of low ATP and higher production of NAD⁺. Lactic acid and NAD⁺ are transported to the extracellular environment, where 585 586 the dinucleotide is consumed by CD38 to trigger intercellular communications and signaling mediated through nucleotides (i.e., cADPR, ADPR, NAADP) and nucleosides (i.e., ADO) (108). 587

588 Patients with severe COVID-19 have been found to have high levels of lactic acid, leading to suppressed proliferation and functions of T lymphocytes (exhaustion). The results are a paresis of 589 cellular and humoral immunities (74). Natural killer (NK) cells exposed to an acidic pH via lactic acid are 590 driven to a state of anergy (19), while acidic conditions inhibit the maturation of DCs and antigen 591 presentation (92). In contrast, myeloid-derived suppressor cells (MDSCs) and regulatory T lymphocytes 592 593 (Treg) are functionally active in acidic environments (91). If these conditions of the immune compartment correlate *in vivo* with the viral infection, it is possible that a dysregulated inflammatory response may 594 595 derive from metabolic acidosis (30). Similar observations have been made in multiple myeloma and severe bacterial sepsis (108, 129). 596

The dysregulated metabolic conditions observed during progression of SARS-CoV-2 infection 597 598 may be brought about by the decay of CD4⁺ Treg cells, which influences hyperinflammation through 599 production of anti-inflammatory ADO (185). ADO has a central role in mediating the pathophysiology of 600 chronic lung diseases (257). The first evidence of a non-canonical adenosinergic pathway involving 601 CD38 activity was described in the human Jurkat T cell line (110). Recent studies have determined that NAD⁺ serves as a precursor to form ADO in the lungs where the dinucleotide is released from human 602 airway epithelial cells and that ectoenzymes (CD38/CD203a/CD73) present in lung cells have the ability 603 to metabolize NAD⁺ to ADO (94). 604

All lung NADase activity was impaired in CD38KO mice as well as in lung membranes suggesting that CD38 is the primary NADase in parenchymal lung cells, whose expression is up-regulated by TNF- α and inhibited by 78c, a pharmacologic blocker of CD38 enzymatic activity (101). The functional impact of the adenosinergic pathway led by CD38 in the lungs may be greater under pathologic conditions, given the overproduction of ADO and the high expression of its receptors in patients with chronic obstructive pulmonary diseases (COPD) (20), confirming the potential therapeutic value of CD38 in lung pathologies [(e.g., acute respiratory distress syndrome (ARDS) and COVID-19].

Generated via a cascade of events triggered by the metabolization of NAD⁺ by CD38 or via ATP degradation (73, 110), ADO regulates innate and adaptive immune responses by stimulating A2A and A2B P1 purinergic receptors (22). ADO ligation to A2A leads to inhibition of the cytolytic activities of
effector T lymphocytes (185) and IFN-γ release by NK cells (168). Moreover, when extracellular ADO
levels are high, ligation of A2B (the low-affinity ADO receptor) influences the antigen-presenting activity
of DCs (64) and activates normal infiltrating cells that block the immune response (such as Tregs,
MDSCs, and macrophages). These effects lead to an established peripheral tolerance (257).
Accordingly, extracellular ADO may act in diseases with essential inflammatory pathognomonic
components (e.g., tumors, COVID-19) as a negative immune checkpoint molecule (108, 213).

At early stages of COVID-19, a severe hypoxia may help induce physiological tissue-protecting 621 622 mechanisms. If left unchecked, they may damage local host tissues. In this sort of scenario, ADO 623 accumulates in the extracellular space of tissues under hypoxic conditions and is able to inhibit the acute inflammatory process via A2A and A2B ADO receptor engagement on immune cells (216, 217). 624 Downstream increase of intracellular cAMP, which in turn inhibits NF-κB-driven inflammation, reduces 625 the damage due to an overactive immune system (186). However, SARS-CoV-2 induces a host pro-626 inflammatory critical life-threatening response, which eventually damages lung epithelial and endothelial 627 cells, impairing the exchange of O2 and CO2 (114). This immune response imbalance induces ARDS, 628 which results in a massive release of inflammatory cytokines or CSS. 629

630 4. Ca^{2+} mediated signals

631 Mobilization of intracellular Ca^{2+} is a universal signaling mechanism to control proliferation, 632 differentiation, transcription, replication and metabolism (37).

633 The endocytic internalization of SARS-CoV-2, the delivery of the viral capsid into the cytoplasm for replication, and the activity of NAD⁺-dependent enzymes, all rely upon Ca²⁺ release from intracellular 634 organelles (103, 106) (Fig. 5). After viral entry, the pathogen-associated molecular patterns (PAMPs) 635 from SARS-CoV-2 are recognized by TLRs (36). The interaction of the TLRs with NF-kB and the adaptor 636 protein MyD88 induces a IFN-1 innate inflammatory response (4). TLRs, MyD88 and NF-kB expression 637 are downregulated by Ang II receptor (AT1R) blockers (ARB), reducing inflammation and protecting lung 638 function (56). In this context, it has been previously established that CD38-mediated Ca²⁺ signaling that 639 640 contributes to Ang II-induced human hepatic fibrosis and increased lung fibrosis in animal models is suppressed by ARB treatment. This adds a reasonable evidence in favor of the therapeutic use of ARB 641 642 in SARS-CoV-2 infection (126, 210).

Activation of CD38 triggers a NAADP/cADPR-Ca²⁺ signaling pathway (Fig. 5). NAADP is formed
 by CD38 catalysis at acidic pH by the exchange of the base NAM of NADP⁺ with NA and localized in EL
 stores (138, 139). In fact, blocking acidic EL stores by inhibiting the vacuolar H⁺-adenosine
 triphosphatase (ATPase) with bafilomycin abrogated NAADP induced Ca²⁺ signaling (86). Downstream
 signaling then initiates DNA transcription for activation of ISGs controlled by the NF-κB transcription
 factor and of the NLRP3 inflammasome (204). The CD38/NAD⁺ pathway is found at the crossroads

between adaptive (i.e., activation of immune cells) and innate immune (i.e., type I IFN-dependent antiviral, the oxidative burst and the proinflammatory responses) defenses. The CD38-induced opening of intracellular Ca²⁺ channels promotes activation of inflammatory and anti-viral processes (174). However, the process of Ca²⁺ mobilization from intracellular stores is exploited by SARS-CoV-2 to trigger the production of highly inflammatory cytokines and profibrotic signals.

This proposed mechanistic model for COVID infection and disease (Fig. 5) focuses on the 654 CD38/NAD⁺ axis, which is at the junction between the oxidative burst (ROS), ISGs, and the 655 hyperinflammatory response. This axis may therefore contribute to viral immunopathology by producing 656 CD38-induced second messengers (cADPR, NAADP and ADPR) with the opening of the RyRs-, TPCs-657 Ca²⁺ channels and through Ca²⁺ influxion via TRPM2. Further, the accumulation of intracellular Ca²⁺ 658 released from EL and ER stores would end with local production of ROS. The outcome would be a 659 contribution of the CD38/NAD⁺ axis and Ca²⁺-mediated signals to the COVID-19 process culminating in 660 661 a cytokine storm syndrome (CSS) and tissue fibrosis.



662

Figure 5. Schematic model showing the potential role of CD38-mediated Ca²⁺ signals in COVID-663 19 pathogenesis. SARS-CoV-2 cell endocytosis depends on the ACE2 catalytic receptor (ACE2R) and 664 proteolytic priming (i.e., TMPRSS2 peptidase) (shown in Fig. 1). Ang II binds to the AT1R to induce 665 activation of either type II- or type III-CD38 catalytic receptor, which in turn stimulates Ca²⁺ release 666 through TPCs and RYRs. Ca²⁺ influx through TRPM2 channels also cooperates to provide a high 667 concentration of Ca²⁺ in the cytosol. The overload of cytosolic Ca²⁺ is involved in the activation of the i) 668 ROS/IFN-type I/ISGs metabolic sequence; ii) NF-kB via PAMPs/TLRs/MvD88-dependent pathway, and 669 iii) NLRP3 inflammasome. This sequence of events is proposed as the likely effects in COVID-19 that 670 culminate in a cytokine storm and multi-organ fibrosis. Pharmacological interventions to control the 671 CD38-dependent NAD⁺ metabolome are being proposed to create hurdles at different steps of SARS-672 CoV-2 infection. ARBs and ACEi i) block (---I) Ang II/AT1R activation, ii) increase expression of ACE2 673

(arrested by viral binding), inducing iii) Ang (1–7) to counterbalance the deleterious pro-inflammatory
effects of Ang II/AT1R (see Fig. 1B). In parallel CD38 activation by Ang II is reduced and consequently
NAD⁺ levels are boosted. Similar effects might be obtained using CD38 inhibitors (CD38inh) or by means
of NAD⁺ precursors supplied. The sACE2 acting as decoy-receptor blocks the viral entry. Therapeutic
checkpoints are depicted as hypothesis-driven, but based on observations in other viral infections,
CD38-related diseases, and preliminary data on COVID-19 (see text).

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681 III. CD38 AND RELATED MOLECULAR PATHWAYS MAY HELP MITIGATE COVID-19 EFFECTS.

682 A. Viral endocytosis and Ca²⁺ mediated signals

During viral Ca²⁺-dependent endocytosis, the S protein is cleaved by TRPMSS2 and by other 683 human enzymes at furin sites [i.e., furin (which is abundant in respiratory tract), and plasmin (involved 684 in fibrinolysis)] to become active to bind to ACE2R. Noteworthy, comorbidities feature elevated levels of 685 the extracellular protease plasmin (119, 224). Following this line, it has been hypothesized that a 686 fibrinolytic inhibition may prove a promising therapeutic target for COVID-19 (119). Virus entry into cells 687 can be also impeded by soluble human recombinant ACE2, which acts as a decoy receptor to hijack 688 689 the virus from the host cellular receptor in very early stages of SARS-CoV-2 infections (167) and, downstream viral infection, by antagonists of Ca²⁺-mediated signals (174). Among these, chloroguine 690 and hydroxychloroquine interfere with Ca²⁺ release from acidic EL (and with the terminal glycosylation 691 692 of ACE2), thus impairing virus-receptor endocytosis in SARS-CoV-2 infection (106, 243, 244).

693 Cell infection generally depends on Ca^{2+} release gated by EL TPCs (29). As with MERS, NAADP-694 dependent Ca^{2+} signaling regulates SAR-CoV-2 translocation from the cell surface to the cytoplasm 695 (99). After CD38 activation, TPCs activity is known to i) alter endolysosomal Ca^{2+} content and pH (24), 696 and to ii) regulate the activity of furin required for proteolytic activation of the viral S protein, fusion 697 activity and cytoplasmic translocation (106, 118). Therefore, direct antagonists of cADPR/NAADP would 698 prevent viral entry in the cell. Indeed, blocking TPCs by the inhibitor tetrandrine strongly inhibited entry 699 of SARS-CoV-2 mediated by S protein (104).

700 **B. CD38 expression and regulation of intracellular Ca²⁺ stores.**

Viral infection awakens different pathways that induce inflammatory conditions. One of these works by activating CD38. TPCs and RYRs are controlled by CD38, and contribute to Ca²⁺ signals responsible for inflammasome activation (24, 29). Thus, inhibition of NAD⁺ and NADP⁺ catabolism mediated by CD38 might interfere with SARS-CoV-2 infection and the inflammatory response.

EL is an acidic compartment ruled by a proton pump. CoV entry is blocked when the pump is inhibited (e.g., by bafilomycin) (85). Given that endosomal acidification depends on proton pump/Ca²⁺ release activities, and that the entry of SARS-CoV-2 is reduced when TPCs are inhibited (104), it would be of considerable interest to know whether modulation of the NAADP-dependent Ca²⁺ signaling generated by CD38/NADase activity interferes with the SARS-CoV-2 pathological process. 710 The inflammatory conditions and the status of macrophages has been monitored using two types 711 of CD38 inhibitor molecules. The first (kuromanin, apigenin and rhein) originates from the flavonoid and anthraquinone families (58, 229), while the second (LX102) is a specifically designed chemical 712 713 compound (254). When applied to explore the functional role of CD38 in macrophages, these treatments 714 suppressed the IL-6 and IL-12 molecules, as well as pathways such as NF-κB, P2Rs, caspase-1 and 715 ERK1/2, all of which are involved in the promotion of inflammation by CD38 (162, 211). Inhibition of CD38 may therefore increase NAD⁺ levels and reduce proinflammatory macrophage polarization, thus 716 717 improving related pathologies (33, 255).

cADPR, ISGs and production of IFN-β are greatly reduced by 8-Bromo-cADPR, a cADPR antagonist, and by kuromanin (58, 203). Both drugs block the release of intracellular Ca²⁺ mediated by the CD38/NAD⁺ axis, thus preventing the onset of a hyperinflammatory condition. Further, kuromanin has an antioxidant function. The anti-oxidative effects of scavenging free radicals may also contribute to warding off inflammation. Inhibition of the enzymatic activities of CD38 may therefore be useful in the design of COVID-19 therapeutics.

Severe lung fibrosis in viral respiratory pathologies might be secondary to high expression of 724 CD38 by endothelial cells. On this line, CD38 has been identified as a key regulator of hepatic stellate 725 726 cells (HSC) activation and reported to increase following the progression of hepatic fibrosis produced 727 as a result of viral infections (161). The fibrotic process in HSC is activated by Ang II and attenuated by 728 AT1R (angiotensin II receptor type 1) blockers (so-called ARB) (224), premises suggestive that the RAS 729 system plays a major role in multi-organ fibrosis (209). The intracellular Ca²⁺ release-dependent pro-730 fibrogenic effects of Ang II/CD38, are further supported by the findings of i) association with increased concentration of TGF-β1 (95) and ii) Ang II-induced overproduction of extracellular matrix proteins (e.g., 731 hvaluronate). The effects on Ca²⁺ elicited by Ang II can be reduced by inhibiting CD38 with 8-Br-cADPR 732 733 or NAM (both cADPR antagonists) or with Ned-19 or dipyridamole (both NAADP competitive antagonists) (126, 148). 734

Furthermore, Ang II-induced Ca²⁺ release is inhibited by staurosporine (a protein kinase C inhibitor) and by scavengers of ROS (179). In addition, NAM prevents tissue damage in animal models with induced lung injury. Indeed, NAM inhibition of CD38 cyclase could attenuate tissue damage induced by Ca2+ signaling (18). In fact, NAM is now included among the treatments against COVID-19 (213, 222).

Lung fibrosis secondary to the SARS-CoV-2 virus is reminiscent of the macrophage activation syndrome (MAS) (42) observed in autoimmune diseases. Examples include systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA), where CD38^{high+} plasma cells play a key role (188). Anti-CD38 mAbs are currently used in the treatment of multiple myeloma (MM), a cancer of the plasma cells, and other hematologic malignancies (156, 160). The results obtained indicate that antibodies also modulate immune cells, including inflammatory monocytes and macrophages. The effects mediated by reacting the catalytic functions and intracellular Ca^{2+} release with CD38 antibodies, still need to be evaluated in COVID-19 patients.

In a variety of cell types, Ca²⁺ homeostasis is regulated by the transcription factor early growth
 response-1 (Egr-1), which promotes Ca²⁺ entry across the PM upon ER-Ca²⁺ store depletion (233).
 Activation of Egr-1, mediated by inhibition of NAD⁺ dependent Sirtuins, is critical for the replication of
 CoV (23). The uptick in the activity of acetylated Egr-1 seen in proinflammatory hyperglycemic
 atherosclerosis (241) highlights an eventual association between the comorbidities and Ca²⁺-dependent
 mechanisms of SARS-CoV-2 infection in determining the aggressiveness of COVID-19 disease.

754 C. CD38 and pregnancy-associated immunosuppression in COVID-19

Pregnant women are more susceptible to respiratory pathogens and severe pneumonia because 755 of the conditions of tolerance established between the immune system of mother and embryo (246). In 756 addition to immunogenetic factors, the ectoenzymatic adenosinergic networks operating in closed 757 environments metabolizes nucleotides (ATP, NAD⁺), providing nucleosides (ADO, INO) with 758 759 immunosuppressive potential (110, 206). The existence of these networks and the contribution of ADOproducing ectoenzymes at the maternal/fetal interface has already been highlighted (27). Accordingly, 760 the impact of COVID-19 infection on pregnant women appears to be less severe or similar to that 761 762 reported for non-pregnant patients who developed COVID-19 pneumonia (253), due to protection of the 763 lungs from CSS brought about by the immune system. Indeed, ADO, acting through the low affinity A2B 764 ADO receptor, stimulates IL-6 and acute-phase inflammatory proteins, such as C-Reactive Protein (CRP) production in macrophages and endothelial cells (154). In fact, reported data show that the 765 766 majority of viral infected pregnant patients had increased IL-6 and CRP (253). Similar trend was reported during the development and progression of MM, where metabolic reprogramming contributes to 767 increasing levels of immunosuppressive ADO (113). This experimental data makes it reasonable to 768 speculate that the induction of ADO within the close placental compartment in gestational patients (27) 769 770 helps mitigate COVID-19 related pneumonia during pregnancy.

771 D. CD38 connections and pharmacological control of COVID-19

Promising therapeutic options include neutralizing antibodies, vaccines, antibody transfer from convalescent-phase plasma, anti-viral proteases, receptor-blocker inhibitors, and drug repurposing (192). Potential therapies (Table 2) include i) small-molecules and drugs as modulators of the CD38/NAD⁺ axis (e.g., CD38 inhibitors, NAM, dexamethasone), ii) soluble factors such as ADO modulators (184) , iii) immunomodulators of CD38 expression (88), as well as iv) immunosuppressive cells (e.g., cytokine-induced killer cells and mesenchymal stem cells) (8, 111).

As previously mentioned, viral infection causes the blocking of surface ACE2 (ACE2R) facilitating the actions of Ang II, thus contributing to COVID-19 pathology (75). It was therefore suggested (225) 780 that an imbalance in the action of ACE1 (that catalyzes Ang II from Ang I) and ACE2 (that catalyzes Ang 781 1-7 from Ang II) may act as primary driver of COVID-19 pathobiology (Fig. 1B). The ACE1/ACE2 782 imbalance occurs due to the viral interference in the ACE2 enzymatic activity, thus i) it enhances Ang II 783 signaling through AT1R associated to harmful effects (vascular and pulmonar tissue injuries), and ii) it 784 reduces Ang 1-7 signaling through its protective MasR (anti-inflammatory, anti-fibrogenic and anti-785 oxidative). Several approaches have been proposed to treat COVID-19 by restoring ACE1/ACE2 steady-state: among these, (i) AT1R antagonists/blockers (ARBs); (ii) ACE1 inhibitors; (iii) agonists of 786 MasR; (iv) recombinant human ACE2 as decoy receptor for the virus, and (v) the development of drugs 787 788 enhancing ACE2 activity. Reducing ACE1/ACE2 imbalance is predicted to blunt COVID-19-associated morbidity and mortality, especially in elderly and vulnerable patients. Importantly, approved direct ARBs 789 (AT1R antagonists) and ACE1 inhibitors (that block the synthesis of Ang II) can be repurposed to test 790 their efficacy in treating COVID-19 (223). Related to this, it was reported that Ang II induces 791 NAADP/cADPR production via CD38, both essential for the entire Ang II-mediated Ca²⁺ signaling. 792 793 Indeed, 8-Br-cADPR antagonizes NAADP production, which was partially blocked by pretreatment with 794 Ned19, a NAADP receptor blocker. Notably, anti-hypertensive ARB-drugs (i.e., losartan) abolished both Ang II-induced NAADP/cADPR production and Ca²⁺ increase (194). 795

The raw material for NAD⁺ biosynthesis, Trp, decreases as a consequence of health disorders (infection, inflammation), thus leading to reduce NAD⁺. Such COVID-19-associated conditions were corrected by prescription of NAD⁺ and/or its precursors (e.g., Trp, NAM, NR, NMN) together with CD38 inhibitor (34). Moreover, clinical trials with ARDS patients, show indeed that NR depresses levels of IL-6, IL-5, IL-2 and TNF- α (55), supporting the view that NAD⁺ boosters might be tested for controlling CSS in COVID-19 patients.

In the human respiratory tract, CD38 expression is regulated by TNF- α , an inflammatory cytokine requiring NF-κB activation, resulting in increased Ca²⁺ responses to agonist corticosteroids (9). Among them, glucocorticoids are used in the management of airway hyperresponsiveness as a result of the negative regulation of genes that promote inflammation or the induction of genes that inhibit inflammation in lung cells (235). The *CD38* promoter region includes NF-κB and glucocorticoid response element (GRE) motifs (158). Thus, CD38 increased expression is down-regulated by dexamethasone through inhibition of NF-κB and its use in COVID-19 has been proposed (136).

The *CD38* gene promoter is also sensitive to vitamins, hormones, cytokines, and different retinoids (158). All-trans retinoic acid (ATRA) is a highly specific inducer of CD38 expression in human myeloid cells mediated through RAR α (54). CD38 expressed by immune cells has been induced by ATRA, which promotes adhesion of cells to endothelium, a feature which is responsible for respiratory distress caused by pulmonary interstitial cell infiltration (the so-called RA syndrome) (83). This event could be the first step towards CSS, which is characteristic of the late phases of COVID-19. In line with this observation, anti-CD38 mAbs specifically block binding of ATRA-treated CD4⁺CD45 T-cells to
endothelium (49), mediated by CD38 interactions between leukocytes and the CD31 antigen present on
the surface of lung endothelial cells.

Extracellular ADO levels governs the switch from the proinflammatory to the suppressive macrophage phenotype (184). This mechanism provides a rationale for targeting the purine metabolism by methotrexate, in order to boost ADO production and reduce the dominance of proinflammatory macrophages. This happens in rheumatic diseases and, potentially, in COVID-19 patients (196, 213).

SARS-CoV-2 grows in the cell, where its ss-RNA is protected from the host's cellular innate immunity (261). To ensures ss-RNA integrity, the viral nsp 10, 14, and 16 are involved in a cap formation (by methylation of the ss-RNA molecule), a process essential for viral replication in host cells (245). Both nsp14 and nsp16 are methylated by S-adenosylmethionine (SAM)-dependent methyltransferase (MTase) enzymes (Fig. 4). As a potential target for antiviral therapy, a complex between SARS-CoV-2 nsp10-nsp16 and a purine adenine (sinefungin) becomes a promising therapeutic approach as a pan-MTase inhibitor (132).

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IV. CD38 IMPACTS BIOLOGICAL MECHANISM(S) THROUGH WHICH SARS-CoV-2 TARGETS ELDERLY PATIENTS WITH ACUTE DISEASE

COVID-19 is a biphasic illness with an innate immune response that transitions into an adaptive immune response except in many elderly patients, who develop severe disease with diffuse lung damage (61). Consequently, the high morbidity in the elderly is a striking feature of COVID-19 (166).

835 A. SARS-CoV-2 cell receptors and senescence

Along with ACE-2, CD26 was also proposed as an endocytic cell receptor for SARS-CoV-2, interacting with the S-protein (239). Both ACE2 and CD26 are associated to senescence: ACE-2 is a known inhibitor of cell proliferation and the RAS system is up-regulated in senescence (125). CD26 is known to be a *bona fide* cell surface marker of senescent cells (155). Similarly, myofibroblasts (which are considered to be senescent and pro-fibrotic cells) also overexpress ACE-2 and CD26 (120, 199). Thus, increased mortality in elderly COVID-19 patients may be related to an increased number of senescent lung cells, which are the main host target for COVID-19 viral infection (26).

843 B. Host defense and maintenance of a balanced inflammatory response in aging

Aging may contribute to the disease scenario through general dysregulation of the immune system, as evinced by increased levels of inflammatory cytokines. Aging is also characterized by increased expression of CD38 in immune cells, resulting in high consumption of the NAD⁺ substrate (31, 255). The consequence is that NAD⁺ depletion may exacerbate the cytokine storm and lead to fatal ARDS, which is most common in older COVID-19 patients (131).

Besides NAD⁺ depletion, ROS detected during hypertension-induced vascular organ damage is 849 850 also associated to aging endothelial cells and fibroblasts (127). This ROS-dependent cell weakness 851 increases with age, and the same is true in older COVID-19 patients (166). ROS damage means that 852 aging cells are unable to express pro-survival antioxidants and anti-inflammatory genes due to 853 dysregulation of the nuclear factor erythroid 2-related factor 2 (NRF2) signaling transcription factor (200, 854 215). In addition, silencing of the antioxidant NRF2 gene results in an increased secretion of proinflammatory cytokines, which mediate CoV-induced CSS (79). Moreover, the cellular levels of the 855 NRF2 protein are down-regulated during RSV infection, promoting ROS damage by triggering NAD⁺-856 857 dependent Sirtuins deacetylation of NRF2 (130). NRF-2 pathway activation is reduced by the CD38 inhibitor kuromanin, which supports the hypothesis of an involvement of the NAD⁺ metabolome during 858 viral infections (69). 859

860 C. The NAD⁺ metabolome in the elderly

The question remains as to why NAD⁺ declines during innate aging and premature aging syndromes (25, 78, 231). The main culprit is CD38, whose expression is physiologically up-regulated during aging (34, 205), particularly in cells targeted by SARS-CoV-2 and expressing high levels of CD38 either at protein or mRNA contents (Fig. 6). These considerations provide support to the pharmacological strategies for reversal of physiological- and pathological-related NAD⁺ depletion and subsequent metabolic dysfunctions (34).

CD38 regulates NAD⁺ homeostasis along with other normal and pathological NAD⁺-dependent cellular processes (1, 159). Among these, CD38 expression is elevated in tissue repair and in fibrotic processes in different organs (209) in a way similar to CD38 up-regulation and NAD⁺ depletion seen in aging. This may suggest intringuing parallels between the biology of aging and fibrogenesis traceable to Ang II/CD38-dependent dysregulation of NAD⁺ homeostasis (126).

Mitochondrial dysfunction occurs during aging due to reduced synthesis of NAD⁺ (25, 63, 90), 872 which could impact macrophage function (165). Interestingly, CD38 is highly expressed in pro-873 inflammatory macrophages (3), and genetic ablation or pharmacological inhibition of CD38 can reverse 874 875 mitochondrial dysfunction and reduce inflammatory cytokines in human monocyte/macrophages and in mice (165, 231). Therefore, it is possible that increased circulating levels of inflammatory factors in an 876 877 imbalanced metabolic cell microenvironment (e.g., in aging, oncogenesis, viral infection) induces CD38 expression, contributing to metabolic dysregulation and in turn promoting the inflammatory function of 878 macrophages in the elderly. 879

NAD⁺ depletion, and the deriving metabolic imbalance (e.g., hypoxia, glycolytic metabolism, and
 increased levels of lactic acid linked to a dysregulation of the immune system) driven by CD38 is
 believed to play a key role in cellular senescence (107). In senescent cells, DNA fragments of nuclear
 origin accumulated in the cytoplasm induce activation of the cGAS-STING cytoplasmic DNA-sensing

machinery (151), with the acquisition of a senescence-associated secretory phenotype (SASP). SASP induces an increase of CD38 expression and subsequent NAD⁺ consumption (31, 32). These SASP⁺senile cells do accumulate in different organs (liver and white adipose tissue) and produce proinflammatory cytokines that promote chronic inflammation and fibrosis (32). The senescent SASP cell is reported to up-regulate CD38 expressed in peripheral macrophages (41); it is thus hypothesized that the accumulation of senescent cells releasing SASP factors increases the activity of CD38, with amplification of cytokine release and NAD⁺ depletion (31, 249).

The senescence/age-related NAD⁺ decline/COVID-19 link may appear paradoxical, since senescent cells do not themselves express high levels of CD38. It may be that the SASP factors upregulate CD38 expression in non-senescent cells (for instance, endothelial cells or M1-macrophages). The SASP circuit might support the relation among cellular senescence, NAD⁺ decline and hyperinflammation, with disruption of cellular NAD⁺ homeostasis and promotion of tissue deterioration (202). In the latter case, senescent cells also secrete proteases, growth factors, and extracellular matrix modifiers, which promote chronic inflammation and fibrosis.

A proteomic database has been compiled of senescence-associated secretomes for aging and several diseases and provides a link between the accumulation of senescent cells and pathological process (10). This data base is expected to shed light on the lesser known aspects of SASP in elderly COVID-19 affected patients and, at the same time, to help disentangle the CD38-dependent mechanisms driving inflammation during SARS-CoV-2 infection in the elderly.

⁹⁰³ CD38 expression is regulated by transcription factor NF- κ B (122), which plays a role in the silent ⁹⁰⁴ inflammation frequently encountered in aging. This suggests that the COVID-19 process may alter the ⁹⁰⁵ NAD⁺/CD38 axis, given that SARS-CoV-2 cell infection dysregulates the NAD⁺ gene set, that includes ⁹⁰⁶ enzymes required for the innate immune response, inducing a severe depletion of NAD⁺ by host cell ⁹⁰⁷ (103).

The reported NAD⁺ attack during aging and the course of COVID-19 involves i) the CD38 NAD⁺-908 glycohydrolase (34, 131), ii) the NAD⁺-dependent Sirtuins, that suppresses both chronic inflammation 909 910 and, by binding to the promoter region of ACE2, viral replication (38), and iii) PARPs, whose transcription is increased in individuals infected with SARS-CoV-2 (103). Other NAD⁺-dependent enzymes are iv) 911 ADP-ribosyltransferases (ARTs) and v) ADP-ribosylhydrolases (ARHs). After ARTs transfer the ADPR 912 unit from NAD⁺ onto an acceptor protein (ADPribosylation), ARHs release the ADPR from the target 913 (128). SARS-CoV-2 possess an ARH involved in cell signaling, gene regulation and apoptosis (128, 914 131), which contributes to the depletion of the already low NAD⁺ levels in aged people. 915

Among other mechanisms involved in age-related declines in NAD⁺ levels, one is the depletion of NAD⁺ precursors (e.g., NMN and NR). Indeed, NAMPT levels also reportedly declining during aging (116). Interestingly, it has been reported that NAMPT and NAD⁺ levels are significantly reduced by TNF-

 α and ROS, impairing the activity of the senescence suppressor Sirtuin1 deacetylase and, therefore, 919 920 contributing to the development of aging-related illnesses and chronic inflammation. Hence, strategies to sustain NAD⁺ biosynthesis might be effective in suppressing physiological and pathological 921 922 inflammation (33, 34). NAD⁺ boosting via dietary NR supplementation (Fig. 4) was shown to improve 923 hepatic fibrosis, while NMN supplementation was shown to reduce pulmonary fibrosis (147). Moreover, 924 78c, a thiazologuin(az)olin(on)e specific CD38 inhibitor, reversed NAD⁺ depletion and reduced the accumulation of inflammatory cells, with a substantial regression of pathological alterations (e.g., fibrotic 925 and inflammatory changes) (101, 231) with therapeutic implications. Indeed, pharmacological 926 927 approaches to boosting NAD⁺ by inhibiting CD38 activity, by NAD⁺ precursor supplementation or by a 928 combination of both, represent potential therapeutic strategies for reversing the consequences of SARS-929 CoV-2 infection.

The main advantages provided by the supply of NAM/NR/NMN regarding NAD⁺ depletion are that: (i) NAM inhibits PARP activity by competing with NAD⁺ for the CD38 active site, thus boosting NAD⁺ homeostatic levels; ii) the increased concentration of NAD⁺ provides the substrate for NAD⁺ kinase, leading to production of NADP⁺, which is a stronger PARP inhibitor (14); (iii) NAM is also able to inhibit Sirtuins (15), thereby replenishing NAD⁺ levels. Overall, the effects of amidated sources for NAD⁺ biogenesis support its use in COVID-19 therapy.

Fibrosis is frequently seen in SARS-CoV-2 inflammation in elderly patients (208, 238). Fibrosis 936 on the basis of persistent DNA damage signaling is reported in SARS-CoV-2 infection (134). Damaged 937 DNA induced PARPs to accumulate free ADPR. Concurrently, NAD⁺ consumption by CD38 generates 938 ADPR that binds to the TRPM2 channel, causing a Ca²⁺ influx across the plasma membrane (228). On 939 the other hand, TPC and RyR, respectively gated by NAADP and cADPR, release intracellular Ca²⁺ 940 from the EL and ER organelles to provide high concentration of Ca²⁺ in the cytosol. The overload of 941 cytosolic Ca²⁺ initiates cell apoptosis along with a cytokine hyperinflammation, potentially causing 942 severe lung failure in COVID-19 patients (Fig. 5). Indeed, the high number of fatalities in elderly COVID-943 19 patients is due to the macrophage overactivation, which leads to a CSS and to lung fibrosis (238). 944

945 Also relevant to the present topic is the recent observation that CD8⁺ tissue-resident memory T cells (Trm) in murine models drive age-associated chronic lung sequelae after viral pneumonia (220). 946 947 The authors found that chronic non-resolving lung pathologies in mice are associated with an accumulation of Trm. However, Trm cells isolated from aged mice display reduced effector functions. 948 The authors demonstrated this is a secondary effect of the lack of a subpopulation expressing molecules 949 involved in TCR signaling. It is reasonable to anticipate that CD38 plays a role in this process. Firstly, 950 the enzymatic roles played by the molecule and derived products may contribute to the effects observed 951 952 (209). Secondly, CD38 is reported as being associated to the TCR/CD3 complex and functionally dependent on it, at least in human models (171, 172). Finally, it would be of interest to investigate the 953

presence of CD203a in the context of the lung environment, before and after viral infection, as a potential
source of immunosuppressive ADO (108).

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959 Figure 6. A. Expression level of CD38 in the principal hematological cell subsets involved in the immune response against viral infections and other diseases. Data were obtained from literature (157, 158), and 960 are a knowledge-based best estimate of the protein expression resulting from evaluation of 961 immunohistochemical staining RNA data and available protein/gene characterization data (N=not 962 detected, L=low expression, M=medium expression, H=high expression). B. CD38 mRNA expression 963 964 levels in hematological tissues and in tissues/organs primarily interested by viral infections and other diseases. Data were obtained from the Human Protein Atlas and are expressed as Consensus 965 Normalized expression (NX), created by combining the data from the three transcriptomics datasets 966 967 (HPA, GTEx and FANTOM5) using the internal normalization pipeline.

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969 V. IMPLICATIONS OF CD38 FOR COVID-19 THERAPY

Despite the well-known multi-faceted biology behind CD38 functions, so far clinical applications in viral infections stay back and still need to be addressed. Moreover, to date no specific drugs and therapeutics are approved by any Regulatory Agencies to prevent or treat SARS-CoV-2 infection. However, the strong groundwork on CD38 provided by theragnostic studies in multiple myeloma and other diseases (Table 1), leave footprints for future research requiring further experimental and preclinical studies (170).

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CD38 IN DISEASE	
DISEASE	POTENTIAL AND THERAPEUTIC APPROACHES
Multiple Myeloma	Elimination of plasma cells through therapeutic anti-CD38 antibodies (ADCC, ADCP, CDC, induction od apoptosis) (52)
Amyloidosis	Elimination of plasma cells (89)
Systemic Lupus Erythematosus (SLE)	Elimination of plasma cells and NK cells (188)
Rheumatoid Arthritis (RA)	Elimination of plasma cells (39)
Systemic Sclerosis (SS)	Mitigation of fibrosis by CD38-targeting of NAD ⁺ metabolism (209)
Chronic active antibody-mediated kidney allograft rejection	Elimination of plasma cells (51)
Neurodegeneration	Age-related modulation of NAD ⁺ metabolism (25)
Еуе	Interaction of neuronal CD38 with the soluble CD31 ligand (97)
Olfactory	Interactions among genes for oxytocin release, oxytocin receptor and CD38 (193)

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982 983 **Table 1**: Potential and therapeutic approaches involving CD38 in diseases. For each disease or organ involved, a potential mechanism of action is suggested. References are included in brackets.

Indeed, the identification of CD38 as a key enzyme involved in NAD⁺ metabolism, cell signaling 985 and immunity strongly suggests its potential as a target in viral pathological conditions. Toward these 986 987 goal, CD38 can be targeted using different pharmacological approaches such as small-molecule inhibitors and enzyme-modulating mAbs. For instance, during viral infection the Ang II dysregulation 988 results in increased signaling through the CD38/NAD⁺-glycohydrolase and purinergic receptors, among 989 others, leading to inflammation, thrombosis, fibrogenic alterations, and organ injury. Accordingly, 990 991 approved drugs that modulate these targets or their ligands (herein discussed) may provide useful 992 therapeutic approaches to blunt multiple aspects of COVID-19 pathology (Table 2). Pharmacological 993 agents used to mitigate the detrimental actions of ACE/Ang II/AT1R axis will not only preserve ACE2 anti-inflammatory functions but also blunt the cytokine storm elicited by SARS-CoV-2 infection. Indeed, 994 ACEi or ARBs leading to reduce Ang II activities, and thus CD38 activation, will help to reduce fibrogenic 995 tissue damages (98). The disruption of Ang II/CD38 axis may also preserves mitochondrial and cellular 996 wellness through AT1R blocking and NAD⁺ boosting. Accordingly, as an AngII/CD38 core-based 997

therapy, a clinical trial has been recently launched recently to test whether (or not) ARBs reduce
respiratory failure in COVID-19 patients (NCT04340557).

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DRUGS	BIOACTIVITY
Inhibitors SARS-CoV-2 endocytosis	
rhACE2 as decoy viral receptor	Blockage of SARS-CoV-2 cell entry (167)
Bafilomycin	Inhibition of Ca ²⁺ release (85)
PanMTase inhibitor Sinefungin	Purine adenine metabolism (132)
Repurposed drugs (HCQ, CQ)	Ca ²⁺ metabolism (244)
Modulators of the RAS system	
AT1R blockers (ARBs)	AT1R antagonists (56)
ACE1 inhibitors (ACEi)	Block the synthesis of Ang II (223)
Agonists of MasR	Activation of Angiotensin protective effects (225)
Drugs enhancing ACE2 Activity	Restoration of ACE1/ACE2 imbalance (223)
Modulators of the CD38/NAD ⁺ axis	
Kuromanin, Apigenin, Rhein, 78c, LX102	CD38/NADase inhibitors (58, 229, 231, 254)
NAD ⁺ , NMN, Vitamin B3 (NAM, NR, NA), Tryp	Restoration of NAD ⁺ levels (15, 32, 33)
Dexamethasone	Downregulation of CD38 expression (136)
Vitamins (Retinoic Acid, D3)	Upregulation of CD38 expression (54, 191)
NAM, 8Br-cADPR	cADPR antagonists (15, 126)
Ned19, Dipyridamole	NAADP antagonists (148, 194)
Soluble Immunomodulators	
Anti-CD38 mAbs (Isatuximab, Daratumumab, MOR202, TAK-079)	Allosteric inhibition of CD38 cyclase activity, cytotoxic effects, clearance of CD38 ⁺ cells (156, 160)
Extracellular ADO	Protection of ARDS patients from hyper-oxigenation damages (40, 62)
Cellular Immunomodulators	
Cytokine-induced killer (CIK) cells	Immunosuppression (8)
Mesenchymal stem cells (MSC)	Immunosuppression (111)

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Table 2: Summary of experimental drugs with potential use in SARS-CoV-2 infection therapy. Each drug is flanked by its mechanism of action controlled by CD38 (details in the text). References are included in brackets.

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The COVID-19 pathological process is associated with increased inflammatory responses, oxidative stress, vascular damage and fibrogenesis. The best clinical strategy for the treatment of COVID-19 patients is known to be a purely supportive care, that includes: i) active hyperoxic ventilation (supplemental oxygen), and measures to prevent infection and worsening of the pathological conditions (232, 247). Unfortunately, this means of oxygenation inhibits the local tissue hypoxia-driven ADO-A2AR-mediated anti-inflammatory protecting mechanism (218), and thereby exacerbates ARDS, a

pathophysiological process that lead to the death of COVID-19 patients (178, 234). As a proof-of-1012 1013 principle, it was reported that a COVID-19 patient with ARDS treated with ADO in high flow 21% O2 aerosol showed an improvement in clinical conditions (62). These effects were confirmed in a pilot trial 1014 1015 with very promising clinical outcomes. Indeed, the pharmacological compensation for the oxygenation-1016 associated loss of the generated extracellular ADO in the lungs of COVID-19 patients was achieved 1017 through intra-tracheal injection or inhalation of synthetic ADO (40). Importantly, the resolution of respiratory failure allowed the authors to concluded that the use of ADO is a valid therapeutic option in 1018 ARDS/COVID-19. 1019

1020 CD38 may be part of the multiple mechanisms explaining the low NAD⁺ levels observed in CD38-1021 related diseases (Table 1). Therefore, inhibition of the CD38 enzymatic activity leading to increased NAD⁺ levels might be of interest for treatment. Unfortunately, the small-molecule inhibitors now available 1022 1023 of CD38 enzymatic activity either have an affinity in the micromolar range (231) or trigger cell cytotoxicity like the therapeutic anti-CD38 mAbs. However, the observed immunosuppressive effects of anti-CD38 1024 1025 mAbs on malignant plasma cells could be useful after regulation of its functional effects. The consequence of a CD38 fine-tuning on the intracellular and extracellular NAD⁺ levels and related 1026 metabolites will help understanding how to modulate CD38 to maximize efficacy and lower potential 1027 1028 adverse events (149).

1029 To explore important aspects of COVID-19 therapeutic drugs, a number of small animal models 1030 (such as mice, hamsters, ferrets) can be used (57, 197). Non-human primate models have also been 1031 explored for COVID-19. Interesting, a characterization of CD38 from cynomolgus macague was reported 1032 and demonstrates genetical, biochemical and immunological similarities of the primate CD38 with the 1033 human protein (71). The study opened new prospects for the pharmacological applications of this 1034 catalytic receptor. Indeed, a current study in cynomolgus macaque has focused on the effect of age on infection with SARS-CoV-2 (173, 256). To facilitate the study of SARS-Co-V2 pathogenesis, and to test 1035 candidate COVID-19 therapeutic agents and drug repurposing, micro-engineered organs-on-chips and 1036 1037 lung organoids as models have been developed (212).

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1039 VI. CONCLUSIONS

The aim of this perspective review is to examine the connections between CD38 and COVID-1041 19. It provides detailed analysis of the mechanisms i) of viral invasion, ii) of viral evasion from innate 1042 and adaptive immune responses, iii) of hyperinflammation associated to metabolic conditions; and 1043 examines the iv) protected immune status during pregnancy and vi) clinical fragility of elderly patients. 1044 To achieve these goals the current review re-analyses hypotheses formulated in the context of RSV 1045 infection by exploiting the results about the role of multi-faceted CD38 in other cellular systems. 1046 Associative basic and clinical research data are herein discussed and integrated with conclusions 1047 reported by others within the field.

We are led to conclude that CD38/NADase is at the centre of a functional axis (i.e., intracellular Ca²⁺ mobilization/IFNs response/ROS burst) exploited by viral infections (e.g., RSV, SARS). Consequently, CD38-induced opening of intracellular Ca²⁺ channels would activate processes able to influence early steps of the disease, but whose persistence and worsening negatively affect the outcome of the COVID-19 disease.

The grounds for this hypothesis are that: i) the substrates of CD38 (i.e., NAD⁺ and NADP⁺) are depleted by viral-induced metabolic rewiring; ii) the products of the enzymatic activities of CD38 (i.e., cADPR/ADPR/NAADP) are involved in an anti-viral and proinflammatory response that may favor the onset of lung immunopathology (i.e., CSS and organ fibrosis). The role of the CD38/NAD⁺ axis at different stages of COVID-19 were also analyzed, along with different therapeutic possibilities. The conclusions are that pathological events of the current pandemic may be mitigated by distinct modulators of the CD38/NAD⁺ axis.

There are still many open questions to be answered concerning i) the impact of the CD38/NAD⁺ axis *in vivo* during SARS-CoV-2 infection; ii) certain mechanisms underlying NAD⁺ involvement during SARS-CoV-2 infection, which remains unclear and requires further research for identifying the precise molecular mechanisms implicated in immunity and metabolic adaptations to SARS-CoV-2 infection, and iii) how to meet the challenge of discovering and developing new therapeutic agents, so critically in demand. However, many of today's findings echo those from past viral infections (e.g., RSV and SARS), thus providing a foothold for dealing with COVID-19.

Of clinical relevance for the future in the strategy to fight COVID-19 is the identification of molecular metabolic pathways generally usurped by the viral pathogen and addressing the evaluation of the impact of agents that selectively target CD38's receptorial and catalytic activities to confirm the potential of CD38 as a novel therapeutic target.

1072 **REFERENCES**

1073

1074 1. **Aksoy P**, **White TA**, **Thompson M**, **Chini EN**. Regulation of intracellular levels of NAD: a novel role 1075 for CD38. *Biochem Biophys Res Commun* 345: 1386–1392, 2006. doi: 10.1016/j.bbrc.2006.05.042.

Alhammad YMO, Kashipathy MM, Roy A, Gagné J-P, McDonald P, Gao P, Nonfoux L, Battaile
 KP, Johnson DK, Holmstrom ED, Poirier GG, Lovell S, Fehr AR. The SARS-CoV-2 conserved
 macrodomain is a mono-ADP-ribosylhydrolase. *J Virol 2020.* doi:10.1128/JVI.01969-20

Amici SA, Young NA, Narvaez-Miranda J, Jablonski KA, Arcos J, Rosas L, Papenfuss TL,
 Torrelles JB, Jarjour WN, Guerau-de-Arellano M. CD38 Is Robustly Induced in Human Macrophages and
 Monocytes in Inflammatory Conditions. *Front Immunol* 9: 1593, 2018. doi: 10.3389/fimmu.2018.01593.

Arancibia SA, Beltrán CJ, Aguirre IM, Silva P, Peralta AL, Malinarich F, Hermoso MA. Toll-like
 receptors are key participants in innate immune responses. *Biol Res* 40: 97–112, 2007. doi: 10.4067/s0716 97602007000200001.

1085 5. **Ausiello CM**, **Urbani F**, **Ia Sala A**, **Funaro A**, **Malavasi F**. CD38 ligation induces discrete cytokine 1086 mRNA expression in human cultured lymphocytes. *Eur J Immunol* 25: 1477–1480, 1995. doi: 1087 10.1002/eji.1830250554.

1088 6. **Badawy AA-B**. Immunotherapy of COVID-19 with poly (ADP-ribose) polymerase inhibitors: starting 1089 with nicotinamide. *Biosci Rep* 40, 2020. doi: 10.1042/BSR20202856.

Balka KR, Louis C, Saunders TL, Smith AM, Calleja DJ, D'Silva DB, Moghaddas F, Tailler M,
 Lawlor KE, Zhan Y, Burns CJ, Wicks IP, Miner JJ, Kile BT, Masters SL, De Nardo D. TBK1 and IKKε Act
 Redundantly to Mediate STING-Induced NF-κB Responses in Myeloid Cells. *Cell Reports* 31: 107492, 2020.
 doi: 10.1016/j.celrep.2020.03.056.

1094 8. **Bamba C**, **Singh SP**, **Choudhury S**. Can mesenchymal stem cell therapy be the interim 1095 management of COVID-19? *Drug Discov Ther* 14: 139–142, 2020. doi: 10.5582/ddt.2020.03032.

1096 9. **Barnes PJ**. Glucocorticosteroids: current and future directions: Glucocorticoids. *British Journal of* 1097 *Pharmacology* 163: 29–43, 2011. doi: 10.1111/j.1476-5381.2010.01199.x.

Basisty N, Kale A, Jeon OH, Kuehnemann C, Payne T, Rao C, Holtz A, Shah S, Sharma V,
 Ferrucci L, Campisi J, Schilling B. A proteomic atlas of senescence-associated secretomes for aging
 biomarker development. *PLOS Biology* 18: e3000599, 2020. doi: 10.1371/journal.pbio.3000599.

1101 11. Baum N, Fliegert R, Bauche A, Hambach J, Menzel S, Haag F, Bannas P, Koch-Nolte F.
1102 Daratumumab and Nanobody-Based Heavy Chain Antibodies Inhibit the ADPR Cyclase but not the NAD+
1103 Hydrolase Activity of CD38-Expressing Multiple Myeloma Cells. *Cancers* 13: 76, 2020. doi:
1104 10.3390/cancers13010076.

Bell TJ, Brand OJ, Morgan DJ, Salek-Ardakani S, Jagger C, Fujimori T, Cholewa L, Tilakaratna
V, Östling J, Thomas M, Day AJ, Snelgrove RJ, Hussell T. Defective lung function following influenza
virus is due to prolonged, reversible hyaluronan synthesis. *Matrix Biology* 80: 14–28, 2019. doi:
10.1016/j.matbio.2018.06.006.

Bestle D, Heindl MR, Limburg H, Van Lam van T, Pilgram O, Moulton H, Stein DA, Hardes K,
Eickmann M, Dolnik O, Rohde C, Klenk H-D, Garten W, Steinmetzer T, Böttcher-Friebertshäuser E.
TMPRSS2 and furin are both essential for proteolytic activation of SARS-CoV-2 in human airway cells. *Life*Sci Alliance 3: e202000786, 2020. doi: 10.26508/lsa.202000786.

1113 14. **Bian C, Zhang C, Luo T, Vyas A, Chen S-H, Liu C, Kassab MA, Yang Y, Kong M, Yu X**. NADP+ 1114 is an endogenous PARP inhibitor in DNA damage response and tumor suppression. *Nat Commun* 10: 693, 1115 2019. doi: 10.1038/s41467-019-08530-5.

1116 15. **Bitterman KJ**, Anderson RM, Cohen HY, Latorre-Esteves M, Sinclair DA. Inhibition of silencing 1117 and accelerated aging by nicotinamide, a putative negative regulator of yeast sir2 and human SIRT1. *J Biol* 1118 Chem 277: 45099–45107, 2002. doi: 10.1074/jbc.M205670200.

1119 16. Blanco-Melo D, Nilsson-Payant BE, Liu W-C, Uhl S, Hoagland D, Møller R, Jordan TX, Oishi K, 1120 Panis M, Sachs D, Wang TT, Schwartz RE, Lim JK, Albrecht RA, tenOever BR. Imbalanced Host 1121 Response to SARS-CoV-2 Drives Development of COVID-19. *Cell* 181: 1036-1045.e9, 2020. doi: 1122 10.1016/j.cell.2020.04.026.

17. Bock KW. Modulation of aryl hydrocarbon receptor (AHR) and the NAD+-consuming enzyme CD38:
Searches of therapeutic options for nonalcoholic fatty liver disease (NAFLD). *Biochem Pharmacol* 175:
1125 113905, 2020. doi: 10.1016/j.bcp.2020.113905.

1126 18. Bogan KL, Brenner C. Nicotinic Acid, Nicotinamide, and Nicotinamide Riboside: A Molecular
1127 Evaluation of NAD ⁺ Precursor Vitamins in Human Nutrition. *Annu Rev Nutr* 28: 115–130, 2008. doi:
1128 10.1146/annurev.nutr.28.061807.155443.

 Brand A, Singer K, Koehl GE, Kolitzus M, Schoenhammer G, Thiel A, Matos C, Bruss C, Klobuch S, Peter K, Kastenberger M, Bogdan C, Schleicher U, Mackensen A, Ullrich E, Fichtner-Feigl S, Kesselring R, Mack M, Ritter U, Schmid M, Blank C, Dettmer K, Oefner PJ, Hoffmann P, Walenta S, Geissler EK, Pouyssegur J, Villunger A, Steven A, Seliger B, Schreml S, Haferkamp S, Kohl E, Karrer S, Berneburg M, Herr W, Mueller-Klieser W, Renner K, Kreutz M. LDHA-Associated Lactic Acid Production Blunts Tumor Immunosurveillance by T and NK Cells. *Cell Metabolism* 24: 657–671, 2016. doi: 10.1016/j.cmet.2016.08.011.

 Brown CD, Benditt JO, Sciurba FC, Lee SM, Criner GJ, Mosenifar Z, Shade DM, Slivka WA,
 Wise RA, National emphysema Treatment Trial Research Group. Exercise testing in severe emphysema: association with quality of life and lung function. *COPD* 5: 117–124, 2008. doi: 10.1080/15412550801941265.

Bruzzone S, Guida L, Zocchi E, Franco L, De Flora A null. Connexin 43 hemi channels mediate
Ca2+-regulated transmembrane NAD+ fluxes in intact cells. *FASEB J* 15: 10–12, 2001. doi: 10.1096/fj.000566fje.

1142 22. **Burnstock G**, **Boeynaems J-M**. Purinergic signalling and immune cells. *Purinergic Signalling* 10: 1143 529–564, 2014. doi: 10.1007/s11302-014-9427-2.

1144 23. **Cai Y**, **Liu Y**, **Zhang X**. Induction of transcription factor Egr-1 gene expression in astrocytoma cells 1145 by Murine coronavirus infection. *Virology* 355: 152–163, 2006. doi: 10.1016/j.virol.2006.07.012.

1146 24. Calcraft PJ, Ruas M, Pan Z, Cheng X, Arredouani A, Hao X, Tang J, Rietdorf K, Teboul L,
1147 Chuang K-T, Lin P, Xiao R, Wang C, Zhu Y, Lin Y, Wyatt CN, Parrington J, Ma J, Evans AM, Galione
1148 A, Zhu MX. NAADP mobilizes calcium from acidic organelles through two-pore channels. *Nature* 459: 596–
1149 600, 2009. doi: 10.1038/nature08030.

1150 25. Camacho-Pereira J, Tarragó MG, Chini CCS, Nin V, Escande C, Warner GM, Puranik AS,
1151 Schoon RA, Reid JM, Galina A, Chini EN. CD38 Dictates Age-Related NAD Decline and Mitochondrial
1152 Dysfunction through an SIRT3-Dependent Mechanism. *Cell Metab* 23: 1127–1139, 2016. doi:
10.1016/j.cmet.2016.05.006.

1154 26. **Campisi J**. Cellular Senescence and Lung Function during Aging. Yin and Yang. *Annals ATS* 13: 1155 S402–S406, 2016. doi: 10.1513/AnnalsATS.201609-703AW.

1156 27. Cecati M, Emanuelli M, Giannubilo SR, Quarona V, Senetta R, Malavasi F, Tranquilli AL,
 1157 Saccucci F. Contribution of adenosine-producing ectoenzymes to the mechanisms underlying the mitigation
 1158 of maternal-fetal conflicts. *J Biol Regul Homeost Agents* 27: 519–529, 2013.

1159 28. Chatterjee S, Daenthanasanmak A, Chakraborty P, Wyatt MW, Dhar P, Selvam SP, Fu J, Zhang
1160 J, Nguyen H, Kang I, Toth K, Al-Homrani M, Husain M, Beeson G, Ball L, Helke K, Husain S, Garrett1161 Mayer E, Hardiman G, Mehrotra M, Nishimura MI, Beeson CC, Bupp MG, Wu J, Ogretmen B, Paulos
1162 CM, Rathmell J, Yu X-Z, Mehrotra S. CD38-NAD+Axis Regulates Immunotherapeutic Anti-Tumor T Cell
1163 Response. *Cell Metabolism* 27: 85-100.e8, 2018. doi: 10.1016/j.cmet.2017.10.006.

1164 29. **Chen X**, **Cao R**, **Zhong W**. Host Calcium Channels and Pumps in Viral Infections. *Cells* 9: 94, 2019. 1165 doi: 10.3390/cells9010094. 30. Chhetri S, Khamis F, Pandak N, Al Khalili H, Said E, Petersen E. A fatal case of COVID-19 due
to metabolic acidosis following dysregulate inflammatory response (cytokine storm). *IDCases* 21: e00829,
2020. doi: 10.1016/j.idcr.2020.e00829.

St. Chini C, Hogan KA, Warner GM, Tarragó MG, Peclat TR, Tchkonia T, Kirkland JL, Chini E. The
NADase CD38 is induced by factors secreted from senescent cells providing a potential link between
senescence and age-related cellular NAD+ decline. *Biochemical and Biophysical Research Communications*513: 486–493, 2019. doi: 10.1016/j.bbrc.2019.03.199.

1173 32. Chini CCS, Peclat TR, Warner GM, Kashyap S, Espindola-Netto JM, de Oliveira GC, Gomez LS,
Hogan KA, Tarragó MG, Puranik AS, Agorrody G, Thompson KL, Dang K, Clarke S, Childs BG,
Kanamori KS, Witte MA, Vidal P, Kirkland AL, De Cecco M, Chellappa K, McReynolds MR, Jankowski
C, Tchkonia T, Kirkland JL, Sedivy JM, van Deursen JM, Baker DJ, van Schooten W, Rabinowitz JD,
Baur JA, Chini EN. CD38 ecto-enzyme in immune cells is induced during aging and regulates NAD+ and
NMN levels. *Nat Metab* 2: 1284–1304, 2020. doi: 10.1038/s42255-020-00298-z.

1179 33. **Chini EN**. CD38 as a regulator of cellular NAD: a novel potential pharmacological target for metabolic 1180 conditions. *Curr Pharm Des* 15: 57–63, 2009. doi: 10.2174/138161209787185788.

1181 34. Chini EN, Chini CCS, Espindola Netto JM, de Oliveira GC, van Schooten W. The Pharmacology
 of CD38/NADase: An Emerging Target in Cancer and Diseases of Aging. *Trends in Pharmacological Sciences* 39: 424–436, 2018. doi: 10.1016/j.tips.2018.02.001.

1184 35. **Chini EN**, **Chini CCS**, **Kato I**, **Takasawa S**, **Okamoto H**. CD38 is the major enzyme responsible for 1185 synthesis of nicotinic acid-adenine dinucleotide phosphate in mammalian tissues. *Biochem J* 362: 125–130, 1186 2002. doi: 10.1042/0264-6021:3620125.

1187 36. **Choudhury A**, **Mukherjee S**. In silico studies on the comparative characterization of the interactions 1188 of SARS-CoV-2 spike glycoprotein with ACE-2 receptor homologs and human TLRs. *J Med Virol.* 92: 2105-1189 2113, 2020. doi: 10.1002/jmv.25987.

1190 37. **Clapham DE**. Calcium Signaling. *Cell* 131: 1047–1058, 2007. doi: 10.1016/j.cell.2007.11.028.

1191 38. Clarke NE, Belyaev ND, Lambert DW, Turner AJ. Epigenetic regulation of angiotensin-converting
enzyme 2 (ACE2) by SIRT1 under conditions of cell energy stress. *Clinical Science* 126: 507–516, 2014. doi:
10.1042/CS20130291.

39. Cole S, Walsh A, Yin X, Wechalekar MD, Smith MD, Proudman SM, Veale DJ, Fearon U, Pitzalis
C, Humby F, Bombardieri M, Axel A, Adams H, Chiu C, Sharp M, Alvarez J, Anderson I, Madakamutil
L, Nagpal S, Guo Y. Integrative analysis reveals CD38 as a therapeutic target for plasma cell-rich predisease and established rheumatoid arthritis and systemic lupus erythematosus. *Arthritis Res Ther* 20: 85,
2018. doi: 10.1186/s13075-018-1578-z.

1199 40. Correale P, Caracciolo M, Bilotta F, Conte M, Cuzzola M, Falcone C, Mangano C, Falzea AC,
1200 Iuliano E, Morabito A, Foti G, Armentano A, Caraglia M, De Lorenzo A, Sitkovsky M, Macheda S.
1201 Therapeutic effects of adenosine in high flow 21% oxygen aereosol in patients with Covid19-pneumonia.
1202 PLoS One 15: e0239692, 2020. doi: 10.1371/journal.pone.0239692.

41. Covarrubias AJ, Kale A, Perrone R, Lopez-Dominguez JA, Pisco AO, Kasler HG, Schmidt MS,
Heckenbach I, Kwok R, Wiley CD, Wong H-S, Gibbs E, Iyer SS, Basisty N, Wu Q, Kim I-J, Silva E,
Vitangcol K, Shin K-O, Lee Y-M, Riley R, Ben-Sahra I, Ott M, Schilling B, Scheibye-Knudsen M,
Ishihara K, Quake SR, Newman J, Brenner C, Campisi J, Verdin E. Senescent cells promote tissue NAD+
decline during ageing via the activation of CD38+ macrophages. *Nat Metab* 2: 1265–1283, 2020. doi:
10.1038/s42255-020-00305-3.

1209 42. **Crayne CB**, **Albeituni S**, **Nichols KE**, **Cron RQ**. The Immunology of Macrophage Activation 1210 Syndrome. *Front Immunol* 10: 119, 2019. doi: 10.3389/fimmu.2019.00119.

43. Dantoft W, Robertson KA, Watkins WJ, Strobl B, Ghazal P. Metabolic Regulators Nampt and Sirt6
Serially Participate in the Macrophage Interferon Antiviral Cascade. *Front Microbiol* 10: 355, 2019. doi:
10.3389/fmicb.2019.00355.

44. Das A, Huang GX, Bonkowski MS, Longchamp A, Li C, Schultz MB, Kim L-J, Osborne B, Joshi
S, Lu Y, Treviño-Villarreal JH, Kang M-J, Hung T, Lee B, Williams EO, Igarashi M, Mitchell JR, Wu LE,
Turner N, Arany Z, Guarente L, Sinclair DA. Impairment of an Endothelial NAD+-H2S Signaling Network
Is a Reversible Cause of Vascular Aging. *Cell* 173: 74-89.e20, 2018. doi: 10.1016/j.cell.2018.02.008.

1218 45. **De Flora A**, **Zocchi E**, **Guida L**, **Franco L**, **Bruzzone S**. Autocrine and paracrine calcium signaling 1219 by the CD38/NAD+/cyclic ADP-ribose system. *Ann N Y Acad Sci* 1028: 176–191, 2004. doi: 1220 10.1196/annals.1322.021.

46. De Flora S, Grassi C, Carati L. Attenuation of influenza-like symptomatology and improvement of
cell-mediated immunity with long-term N-acetylcysteine treatment. *Eur Respir J* 10: 1535–1541, 1997. doi:
10.1183/09031936.97.10071535.

de Graaff PMA, de Jong EC, van Capel TM, van Dijk MEA, Roholl PJM, Boes J, Luytjes W,
 Kimpen JLL, van Bleek GM. Respiratory syncytial virus infection of monocyte-derived dendritic cells
 decreases their capacity to activate CD4 T cells. *J Immunol* 175: 5904–5911, 2005. doi:
 10.4049/jimmunol.175.9.5904.

48. Deaglio S, Dianzani U, Horenstein AL, Fernández JE, van Kooten C, Bragardo M, Funaro A,
Garbarino G, Di Virgilio F, Banchereau J, Malavasi F. Human CD38 ligand. A 120-KDA protein
predominantly expressed on endothelial cells. *J Immunol* 156: 727–734, 1996.

1231 49. Dianzani U, Funaro A, DiFranco D, Garbarino G, Bragardo M, Redoglia V, Buonfiglio D, De
 1232 Monte LB, Pileri A, Malavasi F. Interaction between endothelium and CD4+CD45RA+ lymphocytes. Role
 1233 of the human CD38 molecule. *J Immunol* 153: 952–959, 1994.

1234 50. Diao B, Wang C, Tan Y, Chen X, Liu Y, Ning L, Chen L, Li M, Liu Y, Wang G, Yuan Z, Feng Z,
1235 Zhang Y, Wu Y, Chen Y. Reduction and Functional Exhaustion of T Cells in Patients With Coronavirus
1236 Disease 2019 (COVID-19). *Front Immunol* 11: 827, 2020. doi: 10.3389/fimmu.2020.00827.

1237 51. Doberer K, Kläger J, Gualdoni GA, Mayer KA, Eskandary F, Farkash EA, Agis H, Reiter T, Reindl-Schwaighofer R, Wahrmann M, Cohen G, Haslacher H, Bond G, Simonitsch-Klupp I, Halloran 1238 PF, Böhmig GA. CD38 Antibody Daratumumab for the Treatment of Chronic Active Antibody-mediated 1239 Kidney 1240 Allograft Rejection. Transplantation Publish Ahead of Print, 2020. doi: 10.1097/TP.00000000003247. 1241

1242 52. **Donk NWCJ van de**, **Richardson PG**, **Malavasi F**. CD38 antibodies in multiple myeloma: back to 1243 the future. *Blood* 131: 13–29, 2018. doi: 10.1182/blood-2017-06-740944.

1244 53. Doroftei B, Ilie O-D, Cojocariu R-O, Ciobica A, Maftei R, Grab D, Anton E, McKenna J, Dhunna
 1245 N, Simionescu G. Minireview Exploring the Biological Cycle of Vitamin B3 and Its Influence on Oxidative
 1246 Stress: Further Molecular and Clinical Aspects. *Molecules* 25, 2020. doi: 10.3390/molecules25153323.

1247 54. Drach J, McQueen T, Engel H, Andreeff M, Robertson KA, Collins SJ, Malavasi F, Mehta K.
 1248 Retinoic acid-induced expression of CD38 antigen in myeloid cells is mediated through retinoic acid receptor 1249 alpha. *Cancer Res* 54: 1746–1752, 1994.

1250 55. Elhassan YS, Kluckova K, Fletcher RS, Schmidt MS, Garten A, Doig CL, Cartwright DM, Oakey
1251 L, Burley CV, Jenkinson N, Wilson M, Lucas SJE, Akerman I, Seabright A, Lai Y-C, Tennant DA,
1252 Nightingale P, Wallis GA, Manolopoulos KN, Brenner C, Philp A, Lavery GG. Nicotinamide Riboside
1253 Augments the Aged Human Skeletal Muscle NAD+ Metabolome and Induces Transcriptomic and Anti1254 inflammatory Signatures. *Cell Rep* 28: 1717-1728.e6, 2019. doi: 10.1016/j.celrep.2019.07.043.

1255 56. **Elkahloun AG**, **Saavedra JM**. Candesartan could ameliorate the COVID-19 cytokine storm. *Biomed* 1256 *Pharmacother* 131: 110653, 2020. doi: 10.1016/j.biopha.2020.110653.

1257 57. Enkirch T, von Messling V. Ferret models of viral pathogenesis. *Virology* 479–480: 259–270, 2015.
 1258 doi: 10.1016/j.virol.2015.03.017.

1259 58. Escande C, Nin V, Price NL, Capellini V, Gomes AP, Barbosa MT, O'Neil L, White TA, Sinclair
 1260 DA, Chini EN. Flavonoid Apigenin Is an Inhibitor of the NAD+ase CD38: Implications for Cellular NAD+
 1261 Metabolism, Protein Acetylation, and Treatment of Metabolic Syndrome. *Diabetes* 62: 1084–1093, 2013. doi:

1262 10.2337/db12-1139.

1263 59. Essuman K, Summers DW, Sasaki Y, Mao X, Yim AKY, DiAntonio A, Milbrandt J. TIR Domain
1264 Proteins Are an Ancient Family of NAD+-Consuming Enzymes. *Current Biology* 28: 421-430.e4, 2018. doi:
10.1016/j.cub.2017.12.024.

1266 60. **Estevez B**, **Du X**. New Concepts and Mechanisms of Platelet Activation Signaling. *Physiology* 32: 162–177, 2017. doi: 10.1152/physiol.00020.2016.

1268 61. **Fajgenbaum DC**, **June CH**. Cytokine Storm. *N Engl J Med* 383: 2255–2273, 2020. doi: 1269 10.1056/NEJMra2026131.

Falcone C, Caracciolo M, Correale P, Macheda S, Vadalà EG, La Scala S, Tescione M, Danieli
 R, Ferrarelli A, Tarsitano MG, Romano L, De Lorenzo A. Can Adenosine Fight COVID-19 Acute
 Respiratory Distress Syndrome? *JCM* 9: 3045, 2020. doi: 10.3390/jcm9093045.

1273 63. **Fang EF**, **Bohr VA**. NAD+: The convergence of DNA repair and mitophagy. *Autophagy* 13: 442–443, 2017. doi: 10.1080/15548627.2016.1257467.

1275 64. Fedele G, Di Girolamo M, Recine U, Palazzo R, Urbani F, Horenstein AL, Malavasi F, Ausiello 1276 CM. CD38 Ligation in Peripheral Blood Mononuclear Cells of Myeloma Patients Induces Release of 1277 Protumorigenic IL-6 and Impaired Secretion of IFN γ Cytokines and Proliferation. *Mediators of Inflammation* 1278 2013: 1–7, 2013. doi: 10.1155/2013/564687.

1279 65. Fehr AR, Channappanavar R, Jankevicius G, Fett C, Zhao J, Athmer J, Meyerholz DK, Ahel I,
1280 Perlman S. The Conserved Coronavirus Macrodomain Promotes Virulence and Suppresses the Innate
1281 Immune Response during Severe Acute Respiratory Syndrome Coronavirus Infection. *mBio* 7: e01721-16,
1282 /mbio/7/6/e01721-16.atom, 2016. doi: 10.1128/mBio.01721-16.

1283 66. **Fehr AR**, **Perlman S**. Coronaviruses: An Overview of Their Replication and Pathogenesis. *Methods* 1284 *Mol Biol* 1282: 1–23, 2015. doi: 10.1007/978-1-4939-2438-7_1.

1285 67. **Fehr AR**, **Singh SA**, **Kerr CM**, **Mukai S**, **Higashi H**, **Aikawa M**. The impact of PARPs and ADP-1286 ribosylation on inflammation and host–pathogen interactions. *Genes Dev* 34: 341–359, 2020. doi: 1287 10.1101/gad.334425.119.

1288 68. Ferrari D, Idzko M, Dichmann S, Purlis D, Virchow C, Norgauer J, Chiozzi P, Di Virgilio F,
 1289 Luttmann W. P2 purinergic receptors of human eosinophils: characterization and coupling to oxygen radical
 1290 production. FEBS Lett 486: 217–224, 2000. doi: 10.1016/s0014-5793(00)02306-1.

1291 69. Ferrari D, Speciale A, Cristani M, Fratantonio D, Molonia MS, Ranaldi G, Saija A, Cimino F.
1292 Cyanidin-3-O-glucoside inhibits NF-kB signalling in intestinal epithelial cells exposed to TNF-α and exerts
1293 protective effects via Nrf2 pathway activation. *Toxicol Lett* 264: 51–58, 2016. doi:
1294 10.1016/j.toxlet.2016.10.014.

1295 70. **Ferrario CM**, **Trask AJ**, **Jessup JA**. Advances in biochemical and functional roles of angiotensin-1296 converting enzyme 2 and angiotensin-(1–7) in regulation of cardiovascular function. *American Journal of* 1297 *Physiology-Heart and Circulatory Physiology* 289: H2281–H2290, 2005. doi: 10.1152/ajpheart.00618.2005.

Ferrero E, Orciani M, Vacca P, Ortolan E, Crovella S, Titti F, Saccucci F, Malavasi F.
Characterization and phylogenetic epitope mapping of CD38 ADPR cyclase in the cynomolgus macaque. *BMC Immunol* 5: 21, 2004. doi: 10.1186/1471-2172-5-21.

Ferrero E, Saccucci F, Malavasi F. The Making of a Leukocyte Receptor: Origin, Genes and
Regulation of Human CD38 and Related Molecules. In: *Chemical Immunology and Allergy*, edited by Mehta
K, Malavasi F. KARGER, p. 1–19.

1304 73. **Ferretti E**, **Horenstein AL**, **Canzonetta C**, **Costa F**, **Morandi F**. Canonical and non-canonical 1305 adenosinergic pathways. *Immunology Letters* 205: 25–30, 2019. doi: 10.1016/j.imlet.2018.03.007.

Fischer K, Hoffmann P, Voelkl S, Meidenbauer N, Ammer J, Edinger M, Gottfried E, Schwarz
 S, Rothe G, Hoves S, Renner K, Timischl B, Mackensen A, Kunz-Schughart L, Andreesen R, Krause
 SW, Kreutz M. Inhibitory effect of tumor cell–derived lactic acid on human T cells. *Blood* 109: 3812–3819,

- 1309 2007. doi: 10.1182/blood-2006-07-035972.
- Forrester SJ, Booz GW, Sigmund CD, Coffman TM, Kawai T, Rizzo V, Scalia R, Eguchi S.
 Angiotensin II Signal Transduction: An Update on Mechanisms of Physiology and Pathophysiology. *Physiol Rev* 98: 1627–1738, 2018. doi: 10.1152/physrev.00038.2017.
- Franco R, Lluis C, Canela El, Mallol J, Centelles JJ, Arán JM, Blanco J, Sayós J. Relationships
 Between Metabolic Enzymes and the Nucleoside Transport. In: *Purine and Pyrimidine Metabolism in Man VII*, edited by Harkness RA, Elion GB, Zöllner N. Springer US, p. 395–398.
- Frasca L, Fedele G, Deaglio S, Capuano C, Palazzo R, Vaisitti T, Malavasi F, Ausiello CM. CD38
 orchestrates migration, survival, and Th1 immune response of human mature dendritic cells. *Blood* 107:
 2392–2399, 2006. doi: 10.1182/blood-2005-07-2913.
- 1319 78. Frederick DW, Loro E, Liu L, Davila A, Chellappa K, Silverman IM, Quinn WJ, Gosai SJ, Tichy
 1320 ED, Davis JG, Mourkioti F, Gregory BD, Dellinger RW, Redpath P, Migaud ME, Nakamaru-Ogiso E,
 1321 Rabinowitz JD, Khurana TS, Baur JA. Loss of NAD Homeostasis Leads to Progressive and Reversible
 1322 Degeneration of Skeletal Muscle. *Cell Metab* 24: 269–282, 2016. doi: 10.1016/j.cmet.2016.07.005.
- Fulop GA, Kiss T, Tarantini S, Balasubramanian P, Yabluchanskiy A, Farkas E, Bari F, Ungvari
 Z, Csiszar A. Nrf2 deficiency in aged mice exacerbates cellular senescence promoting cerebrovascular
 inflammation. *GeroScience* 40: 513–521, 2018. doi: 10.1007/s11357-018-0047-6.
- 1326 80. Funaro A, Horenstein AL, Calosso L, Morra M, Tarocco RP, Franco L, De Flora A, Malavasi F.
 1327 Identification and characterization of an active soluble form of human CD38 in normal and pathological fluids.
 1328 Int Immunol 8: 1643–1650, 1996. doi: 10.1093/intimm/8.11.1643.
- 1329 81. Galione A, Evans AM, Ma J, Parrington J, Arredouani A, Cheng X, Zhu MX. The acid test: the
 1330 discovery of two-pore channels (TPCs) as NAADP-gated endolysosomal Ca2+ release channels. *Pflugers* 1331 Arch Eur J Physiol 458: 869–876, 2009. doi: 10.1007/s00424-009-0682-y.
- Ballay N, Anani L, Lopez A, Colombat P, Binet C, Domenech J, Weksler BB, Malavasi F, Herault
 O. The role of platelet/endothelial cell adhesion molecule 1 (CD31) and CD38 antigens in marrow
 microenvironmental retention of acute myelogenous leukemia cells. *Cancer Res* 67: 8624–8632, 2007. doi:
 10.1158/0008-5472.CAN-07-0402.
- 1336 83. Gao Y, Camacho LH, Mehta K. Retinoic acid-induced CD38 antigen promotes leukemia cells
 1337 attachment and interferon-gamma/interleukin-1beta-dependent apoptosis of endothelial cells: implications in
 1338 the etiology of retinoic acid syndrome. *Leuk Res* 31: 455–463, 2007. doi: 10.1016/j.leukres.2006.07.004.
- B4. Garavaglia S, Bruzzone S, Cassani C, Canella L, Allegrone G, Sturla L, Mannino E, Millo E, De
 Flora A, Rizzi M. The high-resolution crystal structure of periplasmic Haemophilus influenzae NAD
 nucleotidase reveals a novel enzymatic function of human CD73 related to NAD metabolism. *Biochem J* 441:
 131–141, 2012. doi: 10.1042/BJ20111263.
- 1343 85. Gerasimenko JV, Charlesworth RM, Sherwood MW, Ferdek PE, Mikoshiba K, Parrington J,
 1344 Petersen OH, Gerasimenko OV. Both RyRs and TPCs are required for NAADP-induced intracellular Ca2+
 1345 release. *Cell Calcium* 58: 237–245, 2015. doi: 10.1016/j.ceca.2015.05.005.
- 1346 86. **Gerasimenko JV**, **Tepikin AV**, **Petersen OH**, **Gerasimenko OV**. Calcium uptake via endocytosis 1347 with rapid release from acidifying endosomes. *Curr Biol* 8: 1335–1338, 1998. doi: 10.1016/s0960-1348 9822(07)00565-9.
- 1349 87. Gharote MA. Role of poly (ADP) ribose polymerase-1 inhibition by nicotinamide as a possible
 1350 additive treatment to modulate host immune response and prevention of cytokine storm in COVID-19. *IJMS*1351 72: 25–28, 2020. doi: 10.25259/IJMS_29_2020.
- 1352 88. Ghobrial I, Cruz CH, Garfall A, Shah N, Munshi N, Kaufman J, Boise LH, Morgan G,
 1353 Adalsteinsson VA, Manier S, Pillai R, Malavasi F, Lonial S. Immunotherapy in Multiple Myeloma:
 1354 Accelerating on the Path to the Patient. *Clin Lymphoma Myeloma Leuk* 19: 332–344, 2019. doi:
 1355 10.1016/j.clml.2019.02.004.

B9. Godara A, Palladini G. Monoclonal Antibody Therapies in Systemic Light-Chain Amyloidosis.
 Hematology/Oncology Clinics of North America 34: 1145–1159, 2020. doi: 10.1016/j.hoc.2020.08.005.

1358 90. Gomes AP, Price NL, Ling AJY, Moslehi JJ, Montgomery MK, Rajman L, White JP, Teodoro
1359 JS, Wrann CD, Hubbard BP, Mercken EM, Palmeira CM, de Cabo R, Rolo AP, Turner N, Bell EL, Sinclair
1360 DA. Declining NAD(+) induces a pseudohypoxic state disrupting nuclear-mitochondrial communication
1361 during aging. *Cell* 155: 1624–1638, 2013. doi: 10.1016/j.cell.2013.11.037.

1362 91. Görgün GT, Whitehill G, Anderson JL, Hideshima T, Maguire C, Laubach J, Raje N, Munshi
1363 NC, Richardson PG, Anderson KC. Tumor-promoting immune-suppressive myeloid-derived suppressor
1364 cells in the multiple myeloma microenvironment in humans. *Blood* 121: 2975–2987, 2013. doi:
1365 10.1182/blood-2012-08-448548.

1366 92. Gottfried E, Kunz-Schughart LA, Ebner S, Mueller-Klieser W, Hoves S, Andreesen R,
1367 Mackensen A, Kreutz M. Tumor-derived lactic acid modulates dendritic cell activation and antigen
1368 expression. *Blood* 107: 2013–2021, 2006. doi: 10.1182/blood-2005-05-1795.

1369 93. Grady SL, Hwang J, Vastag L, Rabinowitz JD, Shenk T. Herpes Simplex Virus 1 Infection Activates
 1370 Poly(ADP-Ribose) Polymerase and Triggers the Degradation of Poly(ADP-Ribose) Glycohydrolase. *J Virol* 1371 86: 8259–8268, 2012. doi: 10.1128/JVI.00495-12.

1372 94. Graeff R, Guedes A, Quintana R, Wendt-Hornickle E, Baldo C, Walseth T, O'Grady S, Kannan
1373 M. Novel Pathway of Adenosine Generation in the Lungs from NAD+: Relevance to Allergic Airway Disease.
1374 Molecules 25: 4966, 2020. doi: 10.3390/molecules25214966.

1375 95. **Gressner AM**, **Weiskirchen R**, **Breitkopf K**, **Dooley S**. Roles of TGF-beta in hepatic fibrosis. *Front* 1376 *Biosci* 7: d793-807, 2002.

1377 96. Grunewald ME, Shaban MG, Mackin SR, Fehr AR, Perlman S. Murine Coronavirus Infection
1378 Activates the Aryl Hydrocarbon Receptor in an Indoleamine 2,3-Dioxygenase-Independent Manner,
1379 Contributing to Cytokine Modulation and Proviral TCDD-Inducible-PARP Expression. *J Virol* 94, 2020. doi:
1380 10.1128/JVI.01743-19.

1381 97. **Guerreiro S**, **Privat A-L**, **Bressac L**, **Toulorge D**. CD38 in Neurodegeneration and 1382 Neuroinflammation. *Cells* 9, 2020. doi: 10.3390/cells9020471.

1383 98. **Gul R**, **Kim U-H**, **Alfadda AA**. Renin-angiotensin system at the interface of COVID-19 infection. *Eur* 1384 *J Pharmacol* 890: 173656, 2021. doi: 10.1016/j.ejphar.2020.173656.

1385 99. **Gunaratne GS**, Yang Y, Li F, Walseth TF, Marchant JS. NAADP-dependent Ca2+ signaling 1386 regulates Middle East respiratory syndrome-coronavirus pseudovirus translocation through the 1387 endolysosomal system. *Cell Calcium* 75: 30–41, 2018. doi: 10.1016/j.ceca.2018.08.003.

1388 100. **Guse AH**, **Diercks B-P**. Integration of nicotinic acid adenine dinucleotide phosphate (NAADP)-1389 dependent calcium signalling: NAADP signalling. *J Physiol* 596: 2735–2743, 2018. doi: 10.1113/JP275974.

1390 101. Haffner CD, Becherer JD, Boros EE, Cadilla R, Carpenter T, Cowan D, Deaton DN, Guo Y,
1391 Harrington W, Henke BR, Jeune MR, Kaldor I, Milliken N, Petrov KG, Preugschat F, Schulte C, Shearer
1392 BG, Shearer T, Smalley TL, Stewart EL, Stuart JD, Ulrich JC. Discovery, Synthesis, and Biological
1393 Evaluation of Thiazoloquin(az)olin(on)es as Potent CD38 Inhibitors. *J Med Chem* 58: 3548–3571, 2015. doi:
1394 10.1021/jm502009h.

Hashimoto T, Perlot T, Rehman A, Trichereau J, Ishiguro H, Paolino M, Sigl V, Hanada T,
 Hanada R, Lipinski S, Wild B, Camargo SMR, Singer D, Richter A, Kuba K, Fukamizu A, Schreiber S,
 Clevers H, Verrey F, Rosenstiel P, Penninger JM. ACE2 links amino acid malnutrition to microbial ecology
 and intestinal inflammation. *Nature* 487: 477–481, 2012. doi: 10.1038/nature11228.

Heer CD, Sanderson DJ, Voth LS, Alhammad YMO, Schmidt MS, Trammell SAJ, Perlman S, 1399 103. 1400 Cohen MS, Fehr AR, Brenner C. Coronavirus infection and PARP expression dysregulate the NAD 1401 Metabolome: an actionable component of innate immunity. J Biol Chem. 2020. 1402 doi:10.1074/jbc.RA120.015138.

1403 104. **Heister PM**, **Poston RN**. Pharmacological hypothesis: TPC2 antagonist tetrandrine as a potential 1404 therapeutic agent for COVID-19. *Pharmacol Res Perspect* 8, 2020. doi: 10.1002/prp2.653.

1405 105. Helmy YA, Fawzy M, Elaswad A, Sobieh A, Kenney SP, Shehata AA. The COVID-19 Pandemic:
1406 A Comprehensive Review of Taxonomy, Genetics, Epidemiology, Diagnosis, Treatment, and Control. *J Clin*1407 *Med* 9, 2020. doi: 10.3390/jcm9041225.

1408 106. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS,
1409 Herrler G, Wu N-H, Nitsche A, Müller MA, Drosten C, Pöhlmann S. SARS-CoV-2 Cell Entry Depends on
1410 ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* 181: 271-280.e8, 2020.
1411 doi: 10.1016/j.cell.2020.02.052.

1412 107. **Hogan KA**, **Chini CCS**, **Chini EN**. The Multi-faceted Ecto-enzyme CD38: Roles in 1413 Immunomodulation, Cancer, Aging, and Metabolic Diseases. *Front Immunol* 10: 1187, 2019. doi: 1414 10.3389/fimmu.2019.01187.

Horenstein AL, Bracci C, Morandi F, Malavasi F. CD38 in Adenosinergic Pathways and Metabolic
 Re-programming in Human Multiple Myeloma Cells: In-tandem Insights From Basic Science to Therapy.
 Front Immunol 10: 760, 2019. doi: 10.3389/fimmu.2019.00760.

Horenstein AL, Chillemi A, Quarona V, Zito A, Roato I, Morandi F, Marimpietri D, Bolzoni M,
Toscani D, Oldham RJ, Cuccioloni M, Sasser AK, Pistoia V, Giuliani N, Malavasi F. NAD⁺-Metabolizing
Ectoenzymes in Remodeling Tumor-Host Interactions: The Human Myeloma Model. *Cells* 4: 520–537, 2015.
doi: 10.3390/cells4030520.

 Horenstein AL, Chillemi A, Zaccarello G, Bruzzone S, Quarona V, Zito A, Serra S, Malavasi F.
 A CD38/CD203a/CD73 ectoenzymatic pathway independent of CD39 drives a novel adenosinergic loop in human T lymphocytes. *Oncolmmunology* 2: e26246, 2013. doi: 10.4161/onci.26246.

Horenstein AL, Chillemi A, Zini R, Quarona V, Bianchi N, Manfredini R, Gambari R, Malavasi F,
 Ferrari D. Cytokine-Induced Killer Cells Express CD39, CD38, CD203a, CD73 Ectoenzymes and P1
 Adenosinergic Receptors. *Front Pharmacol* 9: 196, 2018. doi: 10.3389/fphar.2018.00196.

1428 112. Horenstein AL, Faini AC, Morandi F, Bracci C, Lanza F, Giuliani N, Paulus A, Malavasi F. The
1429 Circular Life of Human CD38: From Basic Science to Clinics and Back. *Molecules* 25: 4844, 2020. doi:
1430 10.3390/molecules25204844.

1431 113. Horenstein AL, Quarona V, Toscani D, Costa F, Chillemi A, Pistoia V, Giuliani N, Malavasi F.
1432 Adenosine Generated in the Bone Marrow Niche Through a CD38-Mediated Pathway Correlates with
1433 Progression of Human Myeloma. *Mol Med* 22: 694–704, 2016. doi: 10.2119/molmed.2016.00198.

Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia
J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q,
Wang J, Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *The Lancet* 395: 497–506, 2020. doi: 10.1016/S0140-6736(20)30183-5.

1438 115. **Idzko M**, **Ferrari D**, **Eltzschig HK**. Nucleotide signalling during inflammation. *Nature* 509: 310–317, 2014. doi: 10.1038/nature13085.

1440 116. **Imai S**. Dissecting systemic control of metabolism and aging in the NAD World: the importance of 1441 SIRT1 and NAMPT-mediated NAD biosynthesis. *FEBS Lett* 585: 1657–1662, 2011. doi: 1442 10.1016/j.febslet.2011.04.060.

1443 117. **Iqbal J**, **Zaidi M**. TNF regulates cellular NAD+ metabolism in primary macrophages. *Biochemical and* 1444 *Biophysical Research Communications* 342: 1312–1318, 2006. doi: 10.1016/j.bbrc.2006.02.109.

1445 118. **Jaimes JA**, **Millet JK**, **Whittaker GR**. Proteolytic Cleavage of the SARS-CoV-2 Spike Protein and 1446 the Role of the Novel S1/S2 Site. *iScience* 23: 101212, 2020. doi: 10.1016/j.isci.2020.101212.

1447 119. **Ji H-L**, **Zhao R**, **Matalon S**, **Matthay MA**. Elevated Plasmin(ogen) as a Common Risk Factor for 1448 COVID-19 Susceptibility. *Physiol Rev* 100: 1065–1075, 2020. doi: 10.1152/physrev.00013.2020.

1449 120. Kam Y-W, Okumura Y, Kido H, Ng LFP, Bruzzone R, Altmeyer R. Cleavage of the SARS

coronavirus spike glycoprotein by airway proteases enhances virus entry into human bronchial epithelial cells
 in vitro. *PLoS One* 4: e7870, 2009. doi: 10.1371/journal.pone.0007870.

1452 121. **Kamel KS**, **Oh MS**, **Halperin ML**. L-lactic acidosis: pathophysiology, classification, and causes; 1453 emphasis on biochemical and metabolic basis. *Kidney Int* 97: 75–88, 2020. doi: 10.1016/j.kint.2019.08.023.

1454 122. Kang B, Tirumurugaan KG, Deshpande DA, Amrani Y, Panettieri RA, Walseth TF, Karman MS, 1455 Kang B, Tirumurugaan KG, Deshpande DA, Amrani Y, Panettieri RA, Walseth TF, Karman MS. 1456 Transcriptional regulation of CD38 expression by tumor necrosis factor- α in human airway smooth muscle 1457 cells: role of NF-κB and sensitivity to glucocorticoids. *FASEB j* 20: 1000–1002, 2006. doi: 10.1096/fj.05-1458 4585fje.

1459 123. Kato K, Nishimasu H, Oikawa D, Hirano S, Hirano H, Kasuya G, Ishitani R, Tokunaga F, Nureki
1460 O. Structural insights into cGAMP degradation by Ecto-nucleotide pyrophosphatase phosphodiesterase 1.
1461 Nat Commun 9: 4424, 2018. doi: 10.1038/s41467-018-06922-7.

1462 124. Katsuyama E, Suarez-Fueyo A, Bradley SJ, Mizui M, Marin AV, Mulki L, Krishfield S, Malavasi
1463 F, Yoon J, Sui SJH, Kyttaris VC, Tsokos GC. The CD38/NAD/SIRTUIN1/EZH2 Axis Mitigates Cytotoxic
1464 CD8 T Cell Function and Identifies Patients with SLE Prone to Infections. *Cell Rep* 30: 112-123.e4, 2020.
1465 doi: 10.1016/j.celrep.2019.12.014.

1466 125. Khemais-Benkhiat S, Idris-Khodja N, Ribeiro TP, Silva GC, Abbas M, Kheloufi M, Lee J-O, Toti
 1467 F, Auger C, Schini-Kerth VB. The Redox-sensitive Induction of the Local Angiotensin System Promotes
 1468 Both Premature and Replicative Endothelial Senescence: Preventive Effect of a Standardized *Crataegus* 1469 Extract. *GERONA* 71: 1581–1590, 2016. doi: 10.1093/gerona/glv213.

1470 126. **Kim S-Y**, **Cho BH**, **Kim U-H**. CD38-mediated Ca2+ signaling contributes to angiotensin II-induced 1471 activation of hepatic stellate cells: attenuation of hepatic fibrosis by CD38 ablation. *J Biol Chem* 285: 576– 1472 582, 2010. doi: 10.1074/jbc.M109.076216.

1473 127. **Kiss T, Balasubramanian P, Valcarcel-Ares MN, Tarantini S, Yabluchanskiy A, Csipo T, Lipecz** 1474 **A, Reglodi D, Zhang XA, Bari F, Farkas E, Csiszar A, Ungvari Z**. Nicotinamide mononucleotide (NMN) 1475 treatment attenuates oxidative stress and rescues angiogenic capacity in aged cerebromicrovascular 1476 endothelial cells: a potential mechanism for the prevention of vascular cognitive impairment. *GeroScience* 1477 41: 619–630, 2019. doi: 10.1007/s11357-019-00074-2.

1478 128. **Koch-Nolte F**. Mammalian ADP-ribosyltransferases and ADP-ribosylhydrolases. *Front Biosci* 1479 Volume: 6716, 2008. doi: 10.2741/3184.

1480 129. **Kogelmann K**, **Jarczak D**, **Scheller M**, **Drüner M**. Hemoadsorption by CytoSorb in septic patients: 1481 a case series. *Crit Care* 21: 74, 2017. doi: 10.1186/s13054-017-1662-9.

1482 130. Komaravelli N, Tian B, Ivanciuc T, Mautemps N, Brasier AR, Garofalo RP, Casola A. Respiratory syncytial virus infection down-regulates antioxidant enzyme expression by triggering deacetylation-1483 degradation 1484 proteasomal of Nrf2. Free Radic Biol Med 88: 391-403. 2015. doi: 10.1016/j.freeradbiomed.2015.05.043. 1485

Kouhpayeh S, Shariati L, Boshtam M, Rahimmanesh I, Mirian M, Zeinalian M, Salari-jazi A,
 Khanahmad N, Damavandi MS, Sadeghi P, Khanahmad H. The Molecular Story of COVID-19; NAD+
 Depletion Addresses All Questions in this Infection. *Preprints* 2020. doi: 10.20944/preprints202003.0346.v1.

1489 132. Krafcikova P, Silhan J, Nencka R, Boura E. Structural analysis of the SARS-CoV-2
1490 methyltransferase complex involved in RNA cap creation bound to sinefungin. *Nat Commun* 11: 3717, 2020.
1491 doi: 10.1038/s41467-020-17495-9.

133. Kuba K, Imai Y, Rao S, Gao H, Guo F, Guan B, Huan Y, Yang P, Zhang Y, Deng W, Bao L, Zhang
B, Liu G, Wang Z, Chappell M, Liu Y, Zheng D, Leibbrandt A, Wada T, Slutsky AS, Liu D, Qin C, Jiang
C, Penninger JM. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus–induced
lung injury. Nat Med 11: 875–879, 2005. doi: 10.1038/nm1267.

1496 134. Kumar V, Agrawal R, Pandey A, Kopf S, Hoeffgen M, Kaymak S, Bandapalli OR, Gorbunova V, 1497 Seluanov A, Mall MA, Herzig S, Nawroth PP. Compromised DNA repair is responsible for diabetes-

- 1498 associated fibrosis. *EMBO J* 39, 2020. doi: 10.15252/embj.2019103477.
- 1499 135. Lange I, Yamamoto S, Partida-Sanchez S, Mori Y, Fleig A, Penner R. TRPM2 functions as a 1500 lysosomal Ca2+-release channel in beta cells. *Sci Signal* 2: ra23, 2009. doi: 10.1126/scisignal.2000278.
- 1501 136. Ledford H. Coronavirus breakthrough: dexamethasone is first drug shown to save lives. *Nature* 582:
 1502 469–469, 2020. doi: 10.1038/d41586-020-01824-5.
- 1503 137. Lee HC. Cyclic ADP-ribose: a calcium mobilizing metabolite of NAD+. *Mol Cell Biochem* 138: 229– 1504 235, 1994. doi: 10.1007/BF00928466.
- 1505 138. Lee HC. Mechanisms of calcium signaling by cyclic ADP-ribose and NAADP. *Physiol Rev* 77: 1133– 1506 1164, 1997. doi: 10.1152/physrev.1997.77.4.1133.
- 1507 139. **Lee HC**. Cyclic ADP-ribose and Nicotinic Acid Adenine Dinucleotide Phosphate (NAADP) as 1508 Messengers for Calcium Mobilization. *J Biol Chem* 287: 31633–31640, 2012. doi: 10.1074/jbc.R112.349464.
- 1509 140. Lee HC, Zhao YJ. Resolving the topological enigma in Ca2+ signaling by cyclic ADP-ribose and 1510 NAADP. *J Biol Chem* 294: 19831–19843, 2019. doi: 10.1074/jbc.REV119.009635.
- 141. Lee S, Paudel O, Jiang Y, Yang X-R, Sham JSK. CD38 Mediates Angiotensin II–Induced
 Intracellular Ca2+ Release in Rat Pulmonary Arterial Smooth Muscle Cells. *Am J Respir Cell Mol Biol* 52:
 332–341, 2015. doi: 10.1165/rcmb.2014-0141OC.
- Lei X, Dong X, Ma R, Wang W, Xiao X, Tian Z, Wang C, Wang Y, Li L, Ren L, Guo F, Zhao Z,
 Zhou Z, Xiang Z, Wang J. Activation and evasion of type I interferon responses by SARS-CoV-2. *Nat Commun* 11: 3810, 2020. doi: 10.1038/s41467-020-17665-9.
- 1517 143. Li X, Geng M, Peng Y, Meng L, Lu S. Molecular immune pathogenesis and diagnosis of COVID-19. 1518 *J Pharm Anal* 10: 102–108, 2020. doi: 10.1016/j.jpha.2020.03.001.
- 1519 144. Liao Y-C, Liang W-G, Chen F-W, Hsu J-H, Yang J-J, Chang M-S. IL-19 Induces Production of IL-1520 6 and TNF- α and Results in Cell Apoptosis Through TNF- α . *J Immunol* 169: 4288–4297, 2002. doi: 1521 10.4049/jimmunol.169.8.4288.
- 1522 145. Liu J, Zhao YJ, Li WH, Hou YN, Li T, Zhao ZY, Fang C, Li SL, Lee HC. Cytosolic interaction of 1523 type III human CD38 with CIB1 modulates cellular cyclic ADP-ribose levels. *Proc Natl Acad Sci USA* 114: 1524 8283–8288, 2017. doi: 10.1073/pnas.1703718114.
- 146. Liu Q, Graeff R, Kriksunov IA, Jiang H, Zhang B, Oppenheimer N, Lin H, Potter BVL, Lee HC,
 Hao Q. Structural Basis for Enzymatic Evolution from a Dedicated ADP-ribosyl Cyclase to a Multifunctional
 NAD Hydrolase. *J Biol Chem* 284: 27637–27645, 2009. doi: 10.1074/jbc.M109.031005.
- 147. Liu T, Rinke AE, Wang J, Phan SH. Cellular NAD+, fibroblast senescence and pulmonary fibrosis.
 FASEB j 34: 1–1, 2020. doi: 10.1096/fasebj.2020.34.s1.02280.
- 148. Liu X, Li Z, Liu S, Sun J, Chen Z, Jiang M, Zhang Q, Wei Y, Wang X, Huang Y-Y, Shi Y, Xu Y,
 Xian H, Bai F, Ou C, Xiong B, Lew AM, Cui J, Fang R, Huang H, Zhao J, Hong X, Zhang Y, Zhou F, Luo
 H-B. Potential therapeutic effects of dipyridamole in the severely ill patients with COVID-19. *Acta Pharm Sin*B 10: 1205–1215, 2020. doi: 10.1016/j.apsb.2020.04.008.
- 1534 149. Liu Y, Clement J, Grant R, Sachdev P, Braidy N. Quantitation of NAD+: Why do we need to 1535 measure it? *Biochimica et Biophysica Acta (BBA) - General Subjects* 1862: 2527–2532, 2018. doi: 1536 10.1016/j.bbagen.2018.07.023.
- 1537 150. Liu Y, Lv J, Liu J, Li M, Xie J, Lv Q, Deng W, Zhou N, Zhou Y, Song J, Wang P, Qin C, Tong W1538 M, Huang B. Mucus production stimulated by IFN-AhR signaling triggers hypoxia of COVID-19. *Cell Res* 30:
 1539 1078–1087, 2020. doi: 10.1038/s41422-020-00435-z.
- 1540 151. **Loring HS**, **Icso JD**, **Nemmara VV**, **Thompson PR**. Initial Kinetic Characterization of Sterile Alpha 1541 and Toll/Interleukin Receptor Motif-Containing Protein 1. *Biochemistry* 59: 933–942, 2020. doi: 1542 10.1021/acs.biochem.9b01078.
- 1543 152. Loring HS, Thompson PR. Emergence of SARM1 as a Potential Therapeutic Target for Wallerian-

1544 type Diseases. *Cell Chem Biol* 27: 1–13, 2020. doi: 10.1016/j.chembiol.2019.11.002.

1545 153. **Loring HS**, **Thompson PR**. A Liquid-to-Solid Phase Transition Enhances the Catalytic Activity of 1546 SARM1. bioRxiv 2020.08.28.272377; doi: https://doi.org/10.1101/2020.08.28.272377

1547 154. Lust JA, Lacy MQ, Zeldenrust SR, Witzig TE, Moon-Tasson LL, Dinarello CA, Donovan KA. 1548 Reduction in C-reactive protein indicates successful targeting of the IL-1/IL-6 axis resulting in improved 1549 survival in early stage multiple myeloma: Targeting IL-1 Induced IL-6 Production. *Am J Hematol* 91: 571– 1550 574, 2016. doi: 10.1002/ajh.24352.

1551 155. Mah W, Jiang G, Olver D, Gallant-Behm C, Wiebe C, Hart DA, Koivisto L, Larjava H, Häkkinen
1552 L. Elevated CD26 Expression by Skin Fibroblasts Distinguishes a Profibrotic Phenotype Involved in Scar
1553 Formation Compared to Gingival Fibroblasts. *Am J Pathol* 187: 1717–1735, 2017. doi:
1554 10.1016/j.ajpath.2017.04.017.

1555 156. **Malavasi F, Chillemi A, Castella B, Schiavoni I, Incarnato D, Oliva S, Horenstein AL**. CD38 and 1556 Antibody Therapy: What Can Basic Science Add? *Blood* 128: SCI-36-SCI-36, 2016. doi: 1557 10.1182/blood.V128.22.SCI-36.SCI-36.

157. Malavasi F, Deaglio S, Ferrero E, Funaro A, Sancho J, Ausiello CM, Ortolan E, Vaisitti T,
Zubiaur M, Fedele G, Aydin S, Tibaldi EV, Durelli I, Lusso R, Cozno F, Horenstein AL. CD38 and CD157
as Receptors of the Immune System: A Bridge Between Innate and Adaptive Immunity. *Mol Med* 12: 334–
341, 2006. doi: 10.2119/2006-00094.Malavasi.

1562 158. Malavasi F, Deaglio S, Funaro A, Ferrero E, Horenstein AL, Ortolan E, Vaisitti T, Aydin S.
1563 Evolution and Function of the ADP Ribosyl Cyclase/CD38 Gene Family in Physiology and Pathology.
1564 Physiological Reviews 88: 841–886, 2008. doi: 10.1152/physrev.00035.2007.

1565 159. Malavasi F, Deaglio S, Zaccarello G, Horenstein AL, Chillemi A, Audrito V, Serra S, Gandione
 1566 M, Zitella A, Tizzani A. The hidden life of NAD+-consuming ectoenzymes in the endocrine system. J
 1567 Molecular Endocrinol 45: 183–191, 2010. doi: 10.1677/JME-10-0082.

160. Manna A, Aulakh S, Jani P, Ahmed S, Akhtar S, Coignet M, Heckman M, Meghji Z, Bhatia K,
Sharma A, Sher T, Alegria V, Malavasi F, Chini EN, Chanan-Khan A, Ailawadhi S, Paulus A. Targeting
CD38 Enhances the Antileukemic Activity of Ibrutinib in Chronic Lymphocytic Leukemia. *Clin Cancer Res* 25:
3974–3985, 2019. doi: 10.1158/1078-0432.CCR-18-3412.

161. March S, Graupera M, Rosa Sarrias M, Lozano F, Pizcueta P, Bosch J, Engel P. Identification
 and Functional Characterization of the Hepatic Stellate Cell CD38 Cell Surface Molecule. *The American Journal of Pathology* 170: 176–187, 2007. doi: 10.2353/ajpath.2007.051212.

162. Matalonga J, Glaria E, Bresque M, Escande C, Carbó JM, Kiefer K, Vicente R, León TE, Beceiro
S, Pascual-García M, Serret J, Sanjurjo L, Morón-Ros S, Riera A, Paytubi S, Juarez A, Sotillo F,
Lindbom L, Caelles C, Sarrias M-R, Sancho J, Castrillo A, Chini EN, Valledor AF. The Nuclear Receptor
LXR Limits Bacterial Infection of Host Macrophages through a Mechanism that Impacts Cellular NAD
Metabolism. *Cell Rep* 18: 1241–1255, 2017. doi: 10.1016/j.celrep.2017.01.007.

1580 163. **Mehraj V**, **Routy J-P**. Tryptophan Catabolism in Chronic Viral Infections: Handling Uninvited Guests. 1581 Int J Tryptophan Res 8: IJTR.S26862, 2015. doi: 10.4137/IJTR.S26862.

1582 164. **Merad M**, **Martin JC**. Pathological inflammation in patients with COVID-19: a key role for monocytes 1583 and macrophages. *Nat Rev Immunol* 20: 355–362, 2020. doi: 10.1038/s41577-020-0331-4.

165. Minhas PS, Liu L, Moon PK, Joshi AU, Dove C, Mhatre S, Contrepois K, Wang Q, Lee BA,
1585 Coronado M, Bernstein D, Snyder MP, Migaud M, Majeti R, Mochly-Rosen D, Rabinowitz JD,
1586 Andreasson KI. Macrophage de novo NAD+ synthesis specifies immune function in aging and inflammation.
1587 Nat Immunol 20: 50–63, 2019. doi: 10.1038/s41590-018-0255-3.

1588 166. Moccia F, Gerbino A, Lionetti V, Miragoli M, Munaron LM, Pagliaro P, Pasqua T, Penna C, 1589 Rocca C, Samaja M, Angelone T. COVID-19-associated cardiovascular morbidity in older adults: a position 1590 paper from the Italian Society of Cardiovascular Researches. *GeroScience* 42: 1021–1049, 2020. doi: 10.1007/s11357-020-00198-w. 167. Monteil V, Kwon H, Prado P, Hagelkrüys A, Wimmer RA, Stahl M, Leopoldi A, Garreta E,
Hurtado del Pozo C, Prosper F, Romero JP, Wirnsberger G, Zhang H, Slutsky AS, Conder R,
Montserrat N, Mirazimi A, Penninger JM. Inhibition of SARS-CoV-2 Infections in Engineered Human
Tissues Using Clinical-Grade Soluble Human ACE2. *Cell* 181: 905-913.e7, 2020. doi:
10.1016/j.cell.2020.04.004.

1597 168. Morandi F, Horenstein AL, Chillemi A, Quarona V, Chiesa S, Imperatori A, Zanellato S, Mortara
 1598 L, Gattorno M, Pistoia V, Malavasi F. CD56brightCD16- NK Cells Produce Adenosine through a CD38 1599 Mediated Pathway and Act as Regulatory Cells Inhibiting Autologous CD4+ T Cell Proliferation. *J Immunol* 1600 195: 965–972, 2015. doi: 10.4049/jimmunol.1500591.

1601 169. **Morandi F**, **Horenstein AL**, **Rizzo R**, **Malavasi F**. The Role of Extracellular Adenosine Generation 1602 in the Development of Autoimmune Diseases. *Mediators Inflamm* 2018: 1–10, 2018. doi: 1603 10.1155/2018/7019398.

1604 170. Morandi F, Marimpietri D, Horenstein AL, Bolzoni M, Toscani D, Costa F, Castella B, Faini AC, Massaia M, Pistoia V, Giuliani N, Malavasi F. Microvesicles released from multiple myeloma cells are 1605 equipped with ectoenzymes belonging to canonical and non-canonical adenosinergic pathways and produce 1606 from ATP and NAD. Oncoimmunology e1458809, 1607 adenosine 7: 2018. doi: 1608 10.1080/2162402X.2018.1458809.

1609 171. **Morra M**, **Zubiaur M**, **Terhorst C**, **Sancho J**, **Malavasi F**. CD38 is functionally dependent on the TCR/CD3 complex in human T cells. *FASEB J* 12: 581–592, 1998. doi: 10.1096/fasebj.12.7.581.

1611 172. **Muñoz P, Mittelbrunn M, de la Fuente H, Pérez-Martínez M, García-Pérez A, Ariza-Veguillas A**, 1612 **Malavasi F, Zubiaur M, Sánchez-Madrid F, Sancho J**. Antigen-induced clustering of surface CD38 and 1613 recruitment of intracellular CD38 to the immunologic synapse. *Blood* 111: 3653–3664, 2008. doi: 1614 10.1182/blood-2007-07-101600.

Munster VJ, Feldmann F, Williamson BN, van Doremalen N, Pérez-Pérez L, Schulz J, Meade White K, Okumura A, Callison J, Brumbaugh B, Avanzato VA, Rosenke R, Hanley PW, Saturday G,
 Scott D, Fischer ER, de Wit E. Respiratory disease and virus shedding in rhesus macaques inoculated with
 SARS-CoV-2. Nature. 2020 Sep;585(7824):268-272. doi: 10.1038/s41586-020-2324-7.

1619 174. Murakami T, Ockinger J, Yu J, Byles V, McColl A, Hofer AM, Horng T. Critical role for calcium
1620 mobilization in activation of the NLRP3 inflammasome. *Proc Nat Acad Sci USA* 109: 11282–11287, 2012.
1621 doi: 10.1073/pnas.1117765109.

1622 175. **Murray MF**, **Nghiem M**, **Srinivasan A**. HIV infection decreases intracellular nicotinamide adenine 1623 dinucleotide [NAD]. *Biochem Biophys Res Commun* 212: 126–131, 1995. doi: 10.1006/bbrc.1995.1945.

1624 176. **Mushtaq M**, **Nam T-S**, **Kim U-H**. Critical role for CD38-mediated Ca2+ signaling in thrombin-induced 1625 procoagulant activity of mouse platelets and hemostasis. *J Biol Chem* 286: 12952–12958, 2011. doi: 1626 10.1074/jbc.M110.207100.

1627 177. **Musso T, Deaglio S, Franco L, Calosso L, Badolato R, Garbarino G, Dianzani U, Malavasi F**. 1628 CD38 expression and functional activities are up-regulated by IFN-gamma on human monocytes and 1629 monocytic cell lines. *J Leukoc Biol* 69: 605–612, 2001.

1630 178. Mutti L, Pentimalli F, Baglio G, Maiorano P, Saladino RE, Correale P, Giordano A. Coronavirus
1631 Disease (Covid-19): What Are We Learning in a Country With High Mortality Rate? *Front Immunol* 11: 1208,
1632 2020. doi: 10.3389/fimmu.2020.01208.

1633 179. Naylor E, Arredouani A, Vasudevan SR, Lewis AM, Parkesh R, Mizote A, Rosen D, Thomas JM,
 1634 Izumi M, Ganesan A, Galione A, Churchill GC. Identification of a chemical probe for NAADP by virtual
 1635 screening. *Nat Chem Biol* 5: 220–226, 2009. doi: 10.1038/nchembio.150.

1636 180. **Newman P, Berndt M**, **Gorski J**, **White G**, **Lyman S**, **Paddock C**, **Muller W**. PECAM-1 (CD31) 1637 cloning and relation to adhesion molecules of the immunoglobulin gene superfamily. *Science* 247: 1219– 1638 1222, 1990. doi: 10.1126/science.1690453.

1639 181. Nieto-Torres JL, Verdiá-Báguena C, Jimenez-Guardeño JM, Regla-Nava JA, Castaño-

Rodriguez C, Fernandez-Delgado R, Torres J, Aguilella VM, Enjuanes L. Severe acute respiratory
 syndrome coronavirus E protein transports calcium ions and activates the NLRP3 inflammasome. *Virology* 485: 330–339, 2015. doi: 10.1016/j.virol.2015.08.010.

1643 182. **Nikiforov A**, **Kulikova V**, **Ziegler M**. The human NAD metabolome: Functions, metabolism and 1644 compartmentalization. *Critical Reviews in Biochemistry and Molecular Biology* 50: 284–297, 2015. doi: 1645 10.3109/10409238.2015.1028612.

1646 183. **Nishina H**, **Inageda K**, **Takahashi K**, **Hoshino S**, **Ikeda K**, **Katada T**. Cell Surface Antigen CD38 1647 Identified as Ecto-Enzyme of NAD Glycohydrolase Has Hyaluronate-Binding Activity. *Biochemical and* 1648 *Biophysical Research Communications* 203: 1318–1323, 1994. doi: 10.1006/bbrc.1994.2326.

184. Ohradanova-Repic A, Machacek C, Charvet C, Lager F, Le Roux D, Platzer R, Leksa V,
Mitulovic G, Burkard TR, Zlabinger GJ, Fischer MB, Feuillet V, Renault G, Blüml S, Benko M, Suchanek
M, Huppa JB, Matsuyama T, Cavaco-Paulo A, Bismuth G, Stockinger H. Extracellular Purine Metabolism
Is the Switchboard of Immunosuppressive Macrophages and a Novel Target to Treat Diseases With
Macrophage Imbalances. *Front Immunol* 9: 852, 2018. doi: 10.3389/fimmu.2018.00852.

1654 185. **Ohta A, Kini R, Ohta A, Subramanian M, Madasu M, Sitkovsky M**. The development and 1655 immunosuppressive functions of CD4+ CD25+ FoxP3+ regulatory T cells are under influence of the 1656 adenosine-A2A adenosine receptor pathway. *Front Immun* 3, 2012. doi: 10.3389/fimmu.2012.00190.

1657 186. **Ohta A**, **Sitkovsky M**. Role of G-protein-coupled adenosine receptors in downregulation of 1658 inflammation and protection from tissue damage. *Nature* 414: 916–920, 2001. doi: 10.1038/414916a.

1659 187. **Onyedibe KI**, **Wang M**, **Sintim HO**. ENPP1, an Old Enzyme with New Functions, and Small Molecule 1660 Inhibitors—A STING in the Tale of ENPP1. *Molecules* 24: 4192, 2019. doi: 10.3390/molecules24224192.

188. Ostendorf L, Burns M, Durek P, Heinz GA, Heinrich F, Garantziotis P, Enghard P, Richter U,
Biesen R, Schneider U, Knebel F, Burmester G, Radbruch A, Mei HE, Mashreghi M-F, Hiepe F,
Alexander T. Targeting CD38 with Daratumumab in Refractory Systemic Lupus Erythematosus. N Engl J
Med 383: 1149–1155, 2020. doi: 10.1056/NEJMoa2023325.

189. Ou X, Liu Y, Lei X, Li P, Mi D, Ren L, Guo L, Guo R, Chen T, Hu J, Xiang Z, Mu Z, Chen X, Chen J, Hu K, Jin Q, Wang J, Qian Z. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nat Commun* 11: 1620, 2020. doi: 10.1038/s41467-020-155629.

1669 190. Partida-Sanchez S, Gasser A, Fliegert R, Siebrands CC, Dammermann W, Shi G, Mousseau
1670 BJ, Sumoza-Toledo A, Bhagat H, Walseth TF, Guse AH, Lund FE. Chemotaxis of Mouse Bone Marrow
1671 Neutrophils and Dendritic Cells Is Controlled by ADP-Ribose, the Major Product Generated by the CD38
1672 Enzyme Reaction. *J Immunol* 179: 7827–7839, 2007. doi: 10.4049/jimmunol.179.11.7827.

1673 191. **Petin K, Weiss R, Müller G, Garten A, Grahnert A, Sack U, Hauschildt S**. NAD metabolites 1674 interfere with proliferation and functional properties of THP-1 cells. *Innate Immun* 25: 280–293, 2019. doi: 10.1177/1753425919844587.

1676 192. **Pillaiyar T**, **Meenakshisundaram S**, **Manickam M**. Recent discovery and development of inhibitors 1677 targeting coronaviruses. *Drug Discovery Today* 25: 668–688, 2020. doi: 10.1016/j.drudis.2020.01.015.

1678 193. Quintana DS, Rokicki J, van der Meer D, Alnæs D, Kaufmann T, Córdova-Palomera A, Dieset
 1679 I, Andreassen OA, Westlye LT. Oxytocin pathway gene networks in the human brain. *Nat Commun* 10:
 1680 668, 2019. doi: 10.1038/s41467-019-08503-8.

1681 194. **Rah S-Y**, **Kim U-H**. Critical Role of CD38 for Generation of Ca2+ Signaling Messengers in 1682 Angiotensin II-Stimulated Kupffer Cells. *Messenger* 1: 77–85, 2012. doi: 10.1166/msr.2012.1004.

1683 195. **Reinherz EL**, **Moretta L**, **Roper M**, **Breard JM**, **Mingari MC**, **Cooper MD**, **Schlossman SF**. Human 1684 T lymphocyte subpopulations defined by Fc receptors and monoclonal antibodies. A comparison. *J Exp Med* 1685 151: 969–974, 1980. doi: 10.1084/jem.151.4.969.

1686 196. Riksen NP, Barrera P, van den Broek PHH, van Riel PLCM, Smits P, Rongen GA. Methotrexate

1687 modulates the kinetics of adenosine in humans in vivo. *Ann Rheum Dis* 65: 465–470, 2006. doi: 10.1136/ard.2005.048637.

197. Rogers TF, Zhao F, Huang D, Beutler N, Burns A, He W-T, Limbo O, Smith C, Song G, Woehl
J, Yang L, Abbott RK, Callaghan S, Garcia E, Hurtado J, Parren M, Peng L, Ramirez S, Ricketts J,
Ricciardi MJ, Rawlings SA, Wu NC, Yuan M, Smith DM, Nemazee D, Teijaro JR, Voss JE, Wilson IA,
Andrabi R, Briney B, Landais E, Sok D, Jardine JG, Burton DR. Isolation of potent SARS-CoV-2
neutralizing antibodies and protection from disease in a small animal model. *Science* 369: 956–963, 2020.
doi: 10.1126/science.abc7520.

198. Sajuthi SP, DeFord P, Jackson ND, Montgomery MT, Everman JL, Rios CL, Pruesse E, Nolin
JD, Plender EG, Wechsler ME, Mak AC, Eng C, Salazar S, Medina V, Wohlford EM, Huntsman S,
Nickerson DA, Germer S, Zody MC, Abecasis G, Kang HM, Rice KM, Kumar R, Oh S, RodriguezSantana J, Burchard EG, Seibold MA. Type 2 and interferon inflammation strongly regulate SARS-CoV-2
related gene expression in the airway epithelium. *Nat Commun* 11: 5139, 2020. doi: 10.1038/s41467-02018781-2.

- 1701 199. **Sargiacomo C**, **Sotgia F**, **Lisanti MP**. COVID-19 and chronological aging: senolytics and other anti-1702 aging drugs for the treatment or prevention of corona virus infection? *Aging* 12: 6511–6517, 2020. doi: 1703 10.18632/aging.103001.
- 1704 200. **Saso L**, **Gürer-Orhan H**, **Stepanić V**. Modulators of Oxidative Stress: Chemical and 1705 Pharmacological Aspects. *Antioxidants* 9: 657, 2020. doi: 10.3390/antiox9080657.
- 1706 201. Savarino A, Bottarel F, Malavasi F, Dianzani U. Role of CD38 in HIV-1 infection: an
 1707 epiphenomenon of T-cell activation or an active player in virus/host interactions?: *AIDS* 14: 1079–1089, 2000.
 1708 doi: 10.1097/00002030-200006160-00004.
- Schafer MJ, White TA, Iijima K, Haak AJ, Ligresti G, Atkinson EJ, Oberg AL, Birch J,
 Salmonowicz H, Zhu Y, Mazula DL, Brooks RW, Fuhrmann-Stroissnigg H, Pirtskhalava T, Prakash YS,
 Tchkonia T, Robbins PD, Aubry MC, Passos JF, Kirkland JL, Tschumperlin DJ, Kita H, LeBrasseur
 NK. Cellular senescence mediates fibrotic pulmonary disease. *Nat Commun* 8: 14532, 2017. doi:
 10.1038/ncomms14532.
- Schiavoni I, Scagnolari C, Horenstein AL, Leone P, Pierangeli A, Malavasi F, Ausiello CM,
 Fedele G. CD38 modulates respiratory syncytial virus-driven proinflammatory processes in human
 monocyte-derived dendritic cells. *Immunology* 154: 122–131, 2018. doi: 10.1111/imm.12873.
- 1717 204. **Schneider WM**, **Chevillotte MD**, **Rice CM**. Interferon-stimulated genes: a complex web of host 1718 defenses. *Annu Rev Immunol* 32: 513–545, 2014. doi: 10.1146/annurev-immunol-032713-120231.
- 1719 205. Schultz MB, Sinclair DA. Why NAD + Declines during Aging: It's Destroyed. *Cell Metabolism* 23:
 1720 965–966, 2016. doi: 10.1016/j.cmet.2016.05.022.
- 1721 206. Serra S, Horenstein AL, Vaisitti T, Brusa D, Rossi D, Laurenti L, D'Arena G, Coscia M, Tripodo
 1722 C, Inghirami G, Robson SC, Gaidano G, Malavasi F, Deaglio S. CD73-generated extracellular adenosine
 1723 in chronic lymphocytic leukemia creates local conditions counteracting drug-induced cell death. *Blood* 118:
 1724 6141–6152, 2011. doi: 10.1182/blood-2011-08-374728.
- 1725 207. Sharma M, Thode T, Weston A, Kaadige MR. Development of Enpp1 Inhibitors as a Strategy to
 1726 Activate Stimulator of Interferon Genes (STING) in Cancers and Other Diseases. *IJCSMB* 5, 2018. doi:
 1727 10.19080/IJCSMB.2018.05.555655.
- Shi B, Bhattacharyya S, Korman B, Marangoni RG, Camp D, Cheresh P, de Oliveira G, Chini E,
 Varga J. Targeting Dysregulated CD38/NAD+ Homeostasis Mitigates Multiple Organ Fibrosis ACR Meeting
 [Abstract] [Online]. *Arthritis Rheumatol* 70, 2018. https://acrabstracts.org/abstract/targeting-dysregulatedcd38-nad-homeostasis-mitigates-multiple-organ-fibrosis/ [26 Oct. 2020].
- Shi B, Wang W, Korman B, Kai L, Wang Q, Wei J, Bale S, Marangoni RG, Bhattacharyya S,
 Miller S, Xu D, Akbarpour M, Cheresh P, Proccissi D, Gursel D, Espindola-Netto JM, Chini CCS, de
 Oliveira GC, Gudjonsson JE, Chini EN, Varga J. Targeting CD38-dependent NAD+ metabolism to mitigate

- 1735 multiple organ fibrosis. *iScience* 24: 101902, 2021. doi: 10.1016/j.isci.2020.101902.
- 1736 210. Shrikrishna D, Astin R, Kemp PR, Hopkinson NS. Renin-angiotensin system blockade: a novel
 1737 therapeutic approach in chronic obstructive pulmonary disease. *Clin Sci (Lond)* 123: 487–498, 2012. doi:
 1738 10.1042/CS20120081.
- Shu B, Feng Y, Gui Y, Lu Q, Wei W, Xue X, Sun X, He W, Yang J, Dai C. Blockade of CD38
 diminishes lipopolysaccharide-induced macrophage classical activation and acute kidney injury involving NFrKB signaling suppression. *Cell Signal* 42: 249–258, 2018. doi: 10.1016/j.cellsig.2017.10.014.
- Si L, Bai H, Rodas M, Cao W, Oh CY, Jiang A, Moller R, Hoagland D, Oishi K, Horiuchi S, Uhl
 S, Blanco-Melo D, Albrecht RA, Liu W-C, Jordan T, Nilsson-Payant BE, Logue J, Haupt R, McGrath M,
 Weston S, Nurani A, Kim SM, Zhu DY, Benam KH, Goyal G, Gilpin SE, Prantil-Baun R, Powers RK,
 Carlson K, Frieman M, tenOever BR, Ingber DE. Human organ chip-enabled pipeline to rapidly repurpose
 therapeutics during viral pandemics. bioRxiv. https://doi. org/10.1101/2020.04.13.039917.
- 1747 213. Sica A, Colombo MP, Trama A, Horn L, Garassino MC, Torri V. Immunometabolic Status of 1748 COVID-19 Cancer Patients. *Physiol Rev* 100: 1839–1850, 2020. doi: 10.1152/physrev.00018.2020.
- 1749 214. **Sica A**, **Mantovani A**. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest* 122: 787– 1750 795, 2012. doi: 10.1172/JCI59643.
- 1751 215. Silva-Palacios A, Königsberg M, Zazueta C. Nrf2 signaling and redox homeostasis in the aging
 1752 heart: A potential target to prevent cardiovascular diseases? *Ageing Res Rev* 26: 81–95, 2016. doi:
 1753 10.1016/j.arr.2015.12.005.
- 1754 216. **Sitkovsky MV**. Use of the A2A adenosine receptor as a physiological immunosuppressor and to 1755 engineer inflammation in vivo. *Biochemical Pharmacology* 65: 493–501, 2003. doi: 10.1016/S0006-1756 2952(02)01548-4.
- 1757 217. Sitkovsky MV, Lukashev D, Apasov S, Kojima H, Koshiba M, Caldwell C, Ohta A, Thiel M. Physiological control of immune response and inflammatory tissue damage by hypoxia-inducible factors and 1758 Immunol 657-682. 1759 adenosine A2A receptors. Annu Rev 22: 2004. doi: 1760 10.1146/annurev.immunol.22.012703.104731.
- 1761 218. **Sitkovsky MV**, **Ohta A**. The "danger" sensors that STOP the immune response: the A2 adenosine 1762 receptors? *Trends Immunol* 26: 299–304, 2005. doi: 10.1016/j.it.2005.04.004.
- 1763 219. **Soares S, Thompson M, White T, Isbell A, Yamasaki M, Prakash Y, Lund FE, Galione A, Chini** 1764 **EN**. NAADP as a second messenger: neither CD38 nor base-exchange reaction are necessary for in vivo 1765 generation of NAADP in myometrial cells. *Am J Physiol Cell Physiol* 292: C227-239, 2007. doi: 1766 10.1152/ajpcell.00638.2005.
- Son YM, Cheon IS, Wu Y, Li C, Wang Z, Gao X, Chen Y, Takahashi Y, Fu Y-X, Dent AL, Kaplan
 MH, Taylor JJ, Cui W, Sun J. Tissue-resident CD4+ T helper cells assist the development of protective
 respiratory B and CD8+ T cell memory responses. *Sci Immunol* 6: eabb6852, 2021. doi:
 10.1126/sciimmunol.abb6852.
- Song E-K, Lee Y-R, Kim Y-R, Yeom J-H, Yoo C-H, Kim H-K, Park H-M, Kang H-S, Kim J-S, Kim
 U-H, Han M-K. NAADP mediates insulin-stimulated glucose uptake and insulin sensitization by PPARγ in adipocytes. *Cell Rep* 2: 1607–1619, 2012. doi: 10.1016/j.celrep.2012.10.018.
- 1774 222. **Song SB**, **Park JS**, **Chung GJ**, **Lee IH**, **Hwang ES**. Diverse therapeutic efficacies and more diverse 1775 mechanisms of nicotinamide. *Metabolomics* 15: 137, 2019. doi: 10.1007/s11306-019-1604-4.
- 1776 223. Sriram K, Insel PA. Risks of ACE Inhibitor and ARB Usage in COVID-19: Evaluating the Evidence.
 1777 *Clin Pharmacol Ther* 108: 236–241, 2020. doi: 10.1002/cpt.1863.
- 1778 224. Sriram K, Insel PA. Inflammation and thrombosis in COVID-19 pathophysiology: Proteinase1779 activated and purinergic receptors as drivers and candidate therapeutic targets. *Physiol Rev.* 2020 Oct 30.
 1780 doi: 10.1152/physrev.00035.2020.
- 1781 225. Sriram K, Insel PA. A hypothesis for pathobiology and treatment of COVID-19: The centrality of

- 1782 ACE1/ACE2 imbalance. *Br J Pharmacol* 177: 4825–4844, 2020. doi: 10.1111/bph.15082.
- 1783 226. **Steardo L**, **Steardo L**, **Zorec R**, **Verkhratsky A**. Neuroinfection may contribute to pathophysiology 1784 and clinical manifestations of COVID-19. *Acta Physiol* 229, 2020. doi: 10.1111/apha.13473.
- 1785 227. **Subbarao K**, **Mahanty S**. Respiratory Virus Infections: Understanding COVID-19. *Immunity* 52: 905– 1786 909, 2020. doi: 10.1016/j.immuni.2020.05.004.
- 1787 228. **Sumoza-Toledo A**, **Penner R**. TRPM2: a multifunctional ion channel for calcium signalling. *J Physiol* 589: 1515–1525, 2011. doi: 10.1113/jphysiol.2010.201855.
- Sun H, Luo G, Chen D, Xiang Z. A Comprehensive and System Review for the Pharmacological
 Mechanism of Action of Rhein, an Active Anthraquinone Ingredient. *Front Pharmacol* 7, 2016. doi:
 10.3389/fphar.2016.00247.
- 1792 230. Tan L, Wang Q, Zhang D, Ding J, Huang Q, Tang Y-Q, Wang Q, Miao H. Lymphopenia predicts
 1793 disease severity of COVID-19: a descriptive and predictive study. *Sig Transduct Target Ther* 5: 33, 2020.
 1794 doi: 10.1038/s41392-020-0148-4.
- 1795 231. Tarragó MG, Chini CCS, Kanamori KS, Warner GM, Caride A, de Oliveira GC, Rud M, Samani
 1796 A, Hein KZ, Huang R, Jurk D, Cho DS, Boslett JJ, Miller JD, Zweier JL, Passos JF, Doles JD, Becherer
 1797 DJ, Chini EN. A Potent and Specific CD38 Inhibitor Ameliorates Age-Related Metabolic Dysfunction by
 1798 Reversing Tissue NAD+ Decline. *Cell Metab* 27: 1081-1095.e10, 2018. doi: 10.1016/j.cmet.2018.03.016.
- 1799 232. **Tay MZ**, **Poh CM**, **Rénia L**, **MacAry PA**, **Ng LFP**. The trinity of COVID-19: immunity, inflammation 1800 and intervention. *Nat Rev Immunol* 20: 363–374, 2020. doi: 10.1038/s41577-020-0311-8.
- 1801 233. Thiel G, Mayer SI, Müller I, Stefano L, Rössler OG. Egr-1—A Ca2+-regulated transcription factor.
 1802 Cell Calcium 47: 397–403, 2010. doi: 10.1016/j.ceca.2010.02.005.
- 1803 234. Thiel M, Chouker A, Ohta A, Jackson E, Caldwell C, Smith P, Lukashev D, Bittmann I, Sitkovsky
 1804 MV. Oxygenation Inhibits the Physiological Tissue-Protecting Mechanism and Thereby Exacerbates Acute
 1805 Inflammatory Lung Injury. *PLoS Biol* 3: e174, 2005. doi: 10.1371/journal.pbio.0030174.
- 1806 235. Tirumurugaan KG, Kang BN, Panettieri RA, Foster DN, Walseth TF, Kannan MS. Regulation of
 1807 the cd38 promoter in human airway smooth muscle cells by TNF-α and dexamethasone. *Respir Res* 9: 26,
 1808 2008. doi: 10.1186/1465-9921-9-26.
- 1809 236. Torti M, Festetics ET, Bertoni A, Sinigaglia F, Balduini C. Thrombin induces the association of
 cyclic ADP-ribose-synthesizing CD38 with the platelet cytoskeleton. *FEBS Letters* 428: 200–204, 1998. doi:
 10.1016/S0014-5793(98)00516-X.
- Vabret N, Britton GJ, Gruber C, Hegde S, Kim J, Kuksin M, Levantovsky R, Malle L, Moreira A, 1812 237. Park MD, Pia L, Risson E, Saffern M, Salomé B, Esai Selvan M, Spindler MP, Tan J, van der Heide V, 1813 1814 Gregory JK, Alexandropoulos K, Bhardwaj N, Brown BD, Greenbaum B, Gümüs ZH, Homann D, Horowitz A, Kamphorst AO, Curotto de Lafaille MA, Mehandru S, Merad M, Samstein RM, Agrawal M, 1815 Aleynick M, Belabed M, Brown M, Casanova-Acebes M, Catalan J, Centa M, Charap A, Chan A, Chen 1816 ST, Chung J, Bozkus CC, Cody E, Cossarini F, Dalla E, Fernandez N, Grout J, Ruan DF, Hamon P, 1817 Humblin E, Jha D, Kodysh J, Leader A, Lin M, Lindblad K, Lozano-Ojalvo D, Lubitz G, Magen A, 1818 Mahmood Z, Martinez-Delgado G, Mateus-Tigue J, Meritt E, Moon C, Noel J, O'Donnell T, Ota M, Plitt 1819 T, Pothula V, Redes J, Reyes Torres I, Roberto M, Sanchez-Paulete AR, Shang J, Schanoski AS, 1820 1821 Suprun M, Tran M, Vaninov N, Wilk CM, Aguirre-Ghiso J, Bogunovic D, Cho J, Faith J, Grasset E, Heeger P, Kenigsberg E, Krammer F, Laserson U. Immunology of COVID-19: Current State of the 1822 Science. Immunity 52: 910–941, 2020. doi: 10.1016/j.immuni.2020.05.002. 1823
- 1824 238. Vaninov N. In the eye of the COVID-19 cytokine storm. *Nat Rev Immunol* 20: 277–277, 2020. doi:
 10.1038/s41577-020-0305-6.
- 1826 239. Vankadari N, Wilce JA. Emerging COVID-19 coronavirus: glycan shield and structure prediction of
 spike glycoprotein and its interaction with human CD26. *Emerging Microbes & Infections* 9: 601–604, 2020.
 1828 doi: 10.1080/22221751.2020.1739565.

1829 240. **Vardhana SA**, **Wolchok JD**. The many faces of the anti-COVID immune response. *Journal of* 1830 *Experimental Medicine* 217: e20200678, 2020. doi: 10.1084/jem.20200678.

Vedantham S, Thiagarajan D, Ananthakrishnan R, Wang L, Rosario R, Zou YS, Goldberg I, Yan
 SF, Schmidt AM, Ramasamy R. Aldose Reductase Drives Hyperacetylation of Egr-1 in Hyperglycemia and
 Consequent Upregulation of Proinflammatory and Prothrombotic Signals. *Diabetes* 63: 761–774, 2014. doi:
 10.2337/db13-0032.

Vollbrecht T, Brackmann H, Henrich N, Roeling J, Seybold U, Bogner JR, Goebel FD, Draenert
 R. Impact of changes in antigen level on CD38/PD-1 co-expression on HIV-specific CD8 T cells in chronic,
 untreated HIV-1 infection. *J Med Virol* 82: 358–370, 2010. doi: 10.1002/jmv.21723.

1838 243. **Walls AC**, **Park Y-J**, **Tortorici MA**, **Wall A**, **McGuire AT**, **Veesler D**. Structure, Function, and 1839 Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell* 181: 281-292.e6, 2020. doi: 1840 10.1016/j.cell.2020.02.058.

1841 244. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, Shi Z, Hu Z, Zhong W, Xiao G. Remdesivir and
1842 chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res* 30: 269–
1843 271, 2020. doi: 10.1038/s41422-020-0282-0.

1844 245. Wang Y, Sun Y, Wu A, Xu S, Pan R, Zeng C, Jin X, Ge X, Shi Z, Ahola T, Chen Y, Guo D.
1845 Coronavirus nsp10/nsp16 Methyltransferase Can Be Targeted by nsp10-Derived Peptide *In Vitro* and *In Vivo*1846 To Reduce Replication and Pathogenesis. *J Virol* 89: 8416–8427, 2015. doi: 10.1128/JVI.00948-15.

1847 246. Wastnedge EAN, Reynolds RM, van Boeckel SR, Stock SJ, Denison FC, Maybin JA, Critchley
1848 HOD. Pregnancy and COVID-19. *Physiol Rev* 101: 303–318, 2021. doi: 10.1152/physrev.00024.2020.

1849 247. Wax RS, Christian MD. Practical recommendations for critical care and anesthesiology teams caring
1850 for novel coronavirus (2019-nCoV) patients. *Can J Anesth/J Can Anesth* 67: 568–576, 2020. doi:
10.1007/s12630-020-01591-x.

1852 248. WHO Solidarity Trial Consortium. Repurposed Antiviral Drugs for Covid-19 — Interim WHO
 1853 Solidarity Trial Results. N Engl J Med 384: 497–511, 2021. doi: 10.1056/NEJMoa2023184.

1854 249. **Wu S**, **Zhang R**. CD38-expressing macrophages drive age-related NAD+ decline. *Nat Metab* 2: 1855 1186–1187, 2020. doi: 10.1038/s42255-020-00292-5.

1856 250. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, Liu S, Zhao P, Liu H, Zhu L, Tai Y, Bai C, Gao
 1857 T, Song J, Xia P, Dong J, Zhao J, Wang F-S. Pathological findings of COVID-19 associated with acute
 1858 respiratory distress syndrome. *Lancet Respir Med* 8: 420–422, 2020. doi: 10.1016/S2213-2600(20)30076-X.

Yamasaki M, Masgrau R, Morgan AJ, Churchill GC, Patel S, Ashcroft SJH, Galione A. Organelle
 selection determines agonist-specific Ca2+ signals in pancreatic acinar and beta cells. *J Biol Chem* 279:
 7234–7240, 2004. doi: 10.1074/jbc.M311088200.

1862 252. **Yang G**. H2S as a potential defense against COVID-19? *Am J Physiol Cell Physiol* 319: C244–C249, 2020. doi: 10.1152/ajpcell.00187.2020.

1864 253. **Yang H**, **Wang C**, **Poon LC**. Novel coronavirus infection and pregnancy. *Ultrasound Obstet Gynecol* 1865 55: 435–437, 2020. doi: 10.1002/uog.22006.

1866 254. Yang L, Li T, Li S, Wu Y, Shi X, Jin H, Liu Z, Zhao Y, Zhang L, Lee HC, Zhang L. Rational Design
and Identification of Small-Molecule Allosteric Inhibitors of CD38. *Chembiochem* 20: 2485–2493, 2019. doi:
10.1002/cbic.201900169.

1869 255. Yarbro JR, Emmons RS, Pence BD. Macrophage Immunometabolism and Inflammaging: Roles of
 1870 Mitochondrial Dysfunction, Cellular Senescence, CD38, and NAD. *Immunometabolism* 2: e200026, 2020.
 1871 doi: 10.20900/immunometab20200026.

1872 256. Yu P, Qi F, Xu Y, Li F, Liu P, Liu J, Bao L, Deng W, Gao H, Xiang Z, Xiao C, Lv Q, Gong S, Liu
1873 J, Song Z, Qu Y, Xue J, Wei Q, Liu M, Wang G, Wang S, Yu H, Liu X, Huang B, Wang W, Zhao L, Wang
1874 H, Ye F, Zhou W, Zhen W, Han J, Wu G, Jin Q, Wang J, Tan W, Qin C. Age-related rhesus macaque
1875 models of COVID-19. *Animal Model Exp Med* 3: 93–97, 2020. doi: 10.1002/ame2.12108.

1876 257. **Zarek PE**, **Powell JD**. Adenosine and anergy. *Autoimmunity* 40: 425–432, 2007. doi: 1877 10.1080/08916930701464939.

1878 258. **Zhao Y**, **Graeff R**, **Lee HC**. Roles of cADPR and NAADP in pancreatic cells. *Acta Biochim Biophys* 1879 *Sin (Shanghai)* 44: 719–729, 2012. doi: 10.1093/abbs/gms044.

259. Zhao ZY, Xie XJ, Li WH, Liu J, Chen Z, Zhang B, Li T, Li SL, Lu JG, Zhang L, Zhang L-H, Xu Z,
 Lee HC, Zhao YJ. A Cell-Permeant Mimetic of NMN Activates SARM1 to Produce Cyclic ADP-Ribose and

1882 Induce Non-apoptotic Cell Death. *iScience* 15: 452–466, 2019. doi: 10.1016/j.isci.2019.05.001.

260. Zhuang M-W, Cheng Y, Zhang J, Jiang X-M, Wang L, Deng J, Wang P-H. Increasing host cellular
 receptor-angiotensin-converting enzyme 2 expression by coronavirus may facilitate 2019-nCoV (or SARS CoV-2) infection. *J Med Virol* 92: 2693–2701, 2020. doi: 10.1002/jmv.26139.

1886 261. **Ziebuhr J**. The Coronavirus Replicase: Insights into a Sophisticated Enzyme Machinery. *Adv Exp* 1887 *Med Biol* 581: 3–11, 2006. doi: 10.1007/978-0-387-33012-9 1.

1889 **FIGURE AND TABLE CAPTIONS:**

1890

Figure 1. Schematic illustration of the SARS-CoV-2 molecular structure and essential 1891 mechanisms of viral infection and outcomes. A) The SARS-CoV-2 genome encodes non-structural 1892 1893 proteins (nsp1-nsp16) (not shown) and four structural proteins: spike (S) glycoprotein, envelope, 1894 membrane, and nucleocapsid phosphoprotein, which together ensure replication of the virus in the host 1895 cell. B) The octapeptide Ang II is originated from the decapeptide Ang I by soluble ACE2 enzymatic 1896 activity. Ang II acts via AT1Rs while Ang (1–7), generated from Ang II by ACE2 carboxypeptidase, acts via the Mas receptor (MasR). SARS-CoV-2 binding to the ACE2 catalytic receptor (ACE2R) enhances 1897 1898 lung inflammation by reducing ACE2 activity and increasing Ang II. Depletion of ACE2 activity decreases the production of Ang 1-7, which has an anti-inflammatory and anti-fibrotic activity. C) SARS-CoV-2 and 1899 1900 RSV preferentially bind to the ACE2R expressed by alveolar epithelial cells and macrophages in the 1901 lower human respiratory tract.

1902

1903 Figure 2. CD38 enzymatic activities. CD38 catalyzes several enzymatic reactions: at neutral pH i) the 1904 conversion of nicotinamide adenine dinucleotide (NAD⁺) into adenosine diphosphate ribose (ADPR) (NAD⁺-glycohydrolase activity); ii) the conversion of NAD⁺ into cyclic ADPR (cADPR) (cyclase activity); 1905 iii) the hydrolysis of cADPR into ADPR (hydrolase activity). At acidic pH, iv) the conversion of NADP⁺, 1906 1907 the phosphorylated equivalent of NAD⁺, into nicotinic acid adenine dinucleotide phosphate (NAADP) (NAADP-synthase activity) in the presence of nicotinic acid (NA) and the degradation of NAADP into 1908 ADPR.P (NAADP-hydrolase activity). All of the reaction products are second messengers involved in 1909 the regulation of cytoplasmic Ca²⁺ fluxes and the generation of immunosuppressive adenosine (see text 1910 and Fig. 3) 1911

1912

1913 Figure 3. Schematic illustration of intracellular signaling mediated by the CD38/NAD⁺ axis. A) The NADPase and NADase enzymes are responsible for the formation of the Ca²⁺-releasing 1914 1915 messengers through the use of phosphorylated (NADP⁺) or non-phosphorylated NAD+, respectively. Second messengers generated as products are: NAADP, cADPR, and ADPR. NAADP-elicited Ca²⁺ is 1916 1917 released from the two-pore channel (TPC) receptor situated in acidic endolysosomes (EL), and cADPR serves as the trigger and booster for Ca²⁺ release via the activation of the ryanodine receptor (RyR), 1918 situated in the endoplasmic reticulum (ER). ADPR elicits Ca²⁺ influx through the transient receptor 1919 melastatin 2 (TRPM2) situated in the plasma membrane (PM). B) ADPR can also be sequentially 1920 metabolized by ectonucleotidases (CD203a/ectonucleotide pyrophosphatase/phosphodiesterase 1 1921 1922 (ENPP1) and CD73/5'-ectonucleotidase (5'eNT) for the formation of extracellular adenosine (ADO).

1923

Figure 4. Pathways for NAD⁺ biogenesis and consumption. Intracellular NAD⁺ is synthesized either 1924 from tryptophan (de novo pathway) or from nicotinamide riboside (NR), nicotinamide (NAM), or nicotinic 1925 1926 acid (NA) (salvage pathways). Once internalized, NAM and NR merge at the step of nicotinamide 1927 mononucleotide (NMN), which is converted into NAD⁺. NA is converted to NA adenine dinucleotide (NAAD), and then to NAD⁺. Depletion of NAD⁺ is associated with enzymatic reactions that take place 1928 intracellularly: CD38/NAD⁺-glycohydrolase, PARPs and Sirtuins. NAD⁺ is also used as a cofactor by S-1929 adenosylmethionine (SAM) for i) the generation of intracellular adenosine from methionine, and ii) the 1930 activity of a viral SAM-dependent Methyl Transferase (MTase) enzyme, composed by the SARS-CoV-1931 2 non-structural proteins (nsp) 14 and 16, active for viral cap formation during viral replication. 1932 1933 Extracellular NAD⁺ is metabolized by CD38, the first enzyme within a purinergic signaling cascade that, 1934 together with CD203 and CD73, generates exogenous adenosine.

1935

Figure 5. Schematic model showing the potential role of CD38-mediated Ca²⁺ signals in COVID-1936 19 pathogenesis. SARS-CoV-2 cell endocytosis depends on the ACE2 catalytic receptor (ACE2R) and 1937 proteolytic priming (i.e., TMPRSS2 peptidase) (shown in Fig. 1). Ang II binds to the AT1R to induce 1938 activation of either type II- or type III-CD38 catalytic receptor, which in turn stimulates Ca²⁺ release 1939 through TPCs and RYRs. Ca²⁺ influx through TRPM2 channels also cooperates to provide a high 1940 concentration of Ca^{2+} in the cytosol. The overload of cytosolic Ca^{2+} is involved in the activation of the i) 1941 1942 ROS/IFN-type I/ISGs metabolic sequence; ii) NF-kB via PAMPs/TLRs/MyD88-dependent pathway, and 1943 iii) NLRP3 inflammasome. This sequence of events is proposed as the likely effects in COVID-19 that 1944 culminate in a cytokine storm and multi-organ fibrosis. Pharmacological interventions to control the CD38-dependent NAD⁺ metabolome are being proposed to create hurdles at different steps of SARS-1945 CoV-2 infection. ARBs and ACEi i) block (---I) Ang II/AT1R activation, ii) increase expression of ACE2 1946 1947 (arrested by viral binding), inducing iii) Ang (1–7) to counterbalance the deleterious pro-inflammatory effects of Ang II/AT1R (see Fig. 1B). In parallel CD38 activation by Ang II is reduced and consequently 1948 1949 NAD⁺ levels are boosted. Similar effects might be obtained using CD38 inhibitors (CD38inh) or by means of NAD⁺ precursors supplied. The sACE2 acting as decoy-receptor blocks the viral entry. Therapeutic 1950 checkpoints are depicted as hypothesis-driven, but based on observations in other viral infections, 1951 CD38-related diseases, and preliminary data on COVID-19 (see text). 1952

1953

Figure 6. A. Expression level of CD38 in the principal hematological cell subsets involved in the immune response against viral infections and other diseases. Data were obtained from literature (157, 158), and are a knowledge-based best estimate of the protein expression resulting from evaluation of immunohistochemical staining RNA data and available protein/gene characterization data (N=not
detected, L=low expression, M=medium expression, H=high expression).
B. CD38 mRNA expression
levels in hematological tissues and in tissues/organs primarily interested by viral infections and other
diseases. Data were obtained from the Human Protein Atlas and are expressed as Consensus
Normalized eXpression (NX), created by combining the data from the three transcriptomics datasets
(HPA, GTEx and FANTOM5) using the internal normalization pipeline.

Table 1: Potential and therapeutic approaches involving CD38 in diseases. For each disease or organ
 involved, a potential mechanism of action is suggested. References are included in brackets.

Table 2: Summary of experimental drugs with potential use in SARS-CoV-2 infection therapy. Each drug
 is flanked by its mechanism of action controlled by CD38 (details in the text). References are included
 in brackets.